

Aberystwyth University

Producing Enhanced Yield and Nutritional Pigmentation in Lollo Rosso Through Manipulating the Irradiance, Duration, and Periodicity of LEDs in the Visible Region of Light

Cammarisano, Laura; Donnison, Iain S.; Robson, Paul R.H.

Published in:
Frontiers in Plant Science

DOI:
[10.3389/fpls.2020.598082](https://doi.org/10.3389/fpls.2020.598082)

Publication date:
2020

Citation for published version (APA):
Cammarisano, L., Donnison, I. S., & Robson, P. R. H. (2020). Producing Enhanced Yield and Nutritional Pigmentation in Lollo Rosso Through Manipulating the Irradiance, Duration, and Periodicity of LEDs in the Visible Region of Light. *Frontiers in Plant Science*, 11, Article 598082. <https://doi.org/10.3389/fpls.2020.598082>

Document License CC BY

General rights

Copyright and moral rights for the publications made accessible in the Aberystwyth Research Portal (the Institutional Repository) are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the Aberystwyth Research Portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the Aberystwyth Research Portal

Take down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

tel: +44 1970 62 2400
email: is@aber.ac.uk



Producing Enhanced Yield and Nutritional Pigmentation in Lollo Rosso Through Manipulating the Irradiance, Duration, and Periodicity of LEDs in the Visible Region of Light

Laura Cammarisano^{1,2*}, Iain S. Donnison¹ and Paul R. H. Robson¹

¹ Institute of Biological, Environmental and Rural Sciences (IBERS), Aberystwyth University, Aberystwyth, United Kingdom,

² Next-Generation Horticultural Systems, Leibniz-Institute of Vegetable and Ornamental Crops (IGZ), Grossbeeren, Germany

OPEN ACCESS

Edited by:

Jason Wargent,
Massey University, New Zealand

Reviewed by:

Cristina Sgheri,
University of Pisa, Italy
Cherubino M. Leonardi,
University of Catania, Italy

*Correspondence:

Laura Cammarisano
cammarisano@igzev.de

Specialty section:

This article was submitted to
Crop and Product Physiology,
a section of the journal
Frontiers in Plant Science

Received: 23 August 2020

Accepted: 23 November 2020

Published: 18 December 2020

Citation:

Cammarisano L, Donnison IS and
Robson PRH (2020) Producing
Enhanced Yield and Nutritional
Pigmentation in Lollo Rosso Through
Manipulating the Irradiance, Duration,
and Periodicity of LEDs in the Visible
Region of Light.
Front. Plant Sci. 11:598082.
doi: 10.3389/fpls.2020.598082

Pigmented food are an important part of the human diet, and anthocyanins have demonstrable protection against tumor production in mouse models and beneficial effects on human liver chemistry. As such, producing pigmented crops is important for a nutritionally diverse diet. Lollo rosso lettuce is a fast-growing pigmented plant, is rich in phenolic compounds, and represents a suitable system to test optimization strategies for yield and anthocyanin production. High-energy UV wavebands are often used to stimulate increased pigmentation; however, we hypothesized that optimizing visible wavebands would deliver both yield and quality improvements. Growing Lollo rosso under irradiances between 5 and 180 W m⁻² using visible waveband LEDs produced 0.4 g fresh weight per W m⁻² in the linear portion of the curve between 5 and 40 W m⁻² and achieved an approximate asymptote of 20 g fresh weight at around 100–120 W m⁻² for yield. Anthocyanin content increased linearly with irradiance. We attempted to optimize the visible wavebands by supplementing half the asymptotic energy for 15 days with supplemental red (R) or blue (B) wavebands in the peaks of photosynthetic activity (430–460 and 630–660 nm). R and B affected rosette morphology with no significant impact on yield, but B significantly increased anthocyanin content by 94% compared to R. We therefore focused on further optimizing B by shortening the daily duration of supplemental B. The minimum B treatment that lacked significant pigment induction was 1 h. We hypothesized that short durations would be more active at different times in the diurnal cycle. Supplemental B was applied for 2 h at four different times. A night-break with B produced the highest yield and anthocyanin content. Our research demonstrates new ways to efficiently use readily available LEDs within the PAR wavebands to increase both yield and crop quality in controlled environment agriculture.

Keywords: LED, blue light, anthocyanin, red light, diel cycle, red lettuce

INTRODUCTION

It is expected that available land per capita to cultivate food will decline due to population growth and climate change. Possible alternatives to increase land use efficiency include using efficient closed-environment agriculture which can produce more crop per unit area (Touliatos et al., 2016). A revolution in lighting allows such systems to utilize efficient LEDs which allow potential control over both irradiance and spectra and, when used in completely enclosed environments, photoperiod (Bantis et al., 2018). Lighting represents a major cost, and therefore, it is important to understand how the increased flexibility achievable in new LED lighting may be best utilized to maximize returns from both crop yield and quality.

Increasing the photon flux density (PFD) results in a linear increase in photosynthetic rate (Kelly et al., 2020) until light exceeds photosynthetic capacity and photosynthesis becomes limited by other factors such as CO₂ (Herron and Mauzerall, 1972; Robinson, 2001). Light use efficiency may also be limited by photoprotective strategies that reduce the risk of photo-oxidative damage by limiting the light interception and absorption and by enhancing dissipative routes (Adams and Demmig-Adams, 1992; Takahashi and Badger, 2011). Such strategies evolved in highly variable environments, and plants can therefore generally tolerate a range of light intensities (Ruban, 2009). Indoor cultivation, where artificial lamps are the sole source of illumination, reduces variability and allows growers to adopt optimal irradiance levels for yield, morphology, and energy costs (Kozai, 2013). Growing plants under optimized light conditions can lead to high yield and high-quality produce (Ouzounis et al., 2015). The exact light recipe may require a balance of light that is optimally used for photosynthesis and that also induces other characteristics of commercial value such as pigmentation and morphological changes that under natural conditions are responses to light stress.

Growing plants under monochromatic R or B appears to be unsuitable for most plants tested so far; monochromatic B results in a decline in photosynthetic activity (Lichtenthaler et al., 1980) and monochromatic R in the deleterious red light syndrome (Matsuda et al., 2004). A combination of R and B radiation, with notable variations in the importance of the ratio between R and B, represents a suitable and efficient light spectrum for crop growth and development (Kim et al., 2004; Izzo et al., 2019). However, a broader spectrum may be more beneficial for crop growth (Pennisi et al., 2019) and addition of green light to the growth spectrum enhances plant growth and development (Johkan et al., 2012) and increases light to the lower plant canopy (Terashima et al., 2009). Replacing a part of the broad-spectrum background with an appropriate fraction of B or R light may increase biomass accumulation (Kaiser et al., 2019), produce regular crop morphological and physiological characteristics, and enhance crop quality (Li and Kubota, 2009).

The types of crops predominantly grown in controlled environments under LEDs have tended to be limited to leafy greens and micro herbs (Benke and Tomkins, 2017). Many studies have focused on lettuce as an important global crop that responds well to light treatment (Son and Oh, 2013).

Lollo rosso lettuce is rapid growing and red pigmented, the major pigment being the anthocyanidin cyanidin (Ferrerres et al., 1997). Anthocyanin showed anti-carcinogenic activity in cell culture models and in animal model tumor systems (Wang and Stoner, 2008), and some pigments can protect lipoproteins and vascular cells from oxidation which is the widely accepted theory for the genesis of atherosclerosis (Wilcox et al., 2003). The importance of pigments has led to the general advice to eat a more varied and colorful diet (WHO-FAO, 2004). Furthermore, anthocyanin accumulation is stimulated by both light intensity (Stutte and Edney, 2009) and quality (Zhang et al., 2018) in red lettuce. As such, Lollo rosso represents a suitable system for cultivation in controlled environment agriculture for studies of both yield and important nutritional quality.

Reports of optimal irradiance for indoor cultivation of lettuce vary between 100 and 600 $\mu\text{mol m}^{-2} \text{s}^{-1}$ (Hee and Beom, 2001; Ilieva et al., 2010; Fu et al., 2012; Kang et al., 2013). The ratio of R to B light affects growth in lettuce (Chen et al., 2016; Clavijo-Herrera et al., 2018; Naznin et al., 2019), and there is a clear interaction between the amount of R, B, and W (Son et al., 2016). In addition to the impacts of spectrum, the timing and duration of light treatments may also be significant and although circadian biology is well researched in plant sciences its application to closed environment agriculture is sparse. Many physiological plant processes change over the day in response to environmental signals; others instead follow specific cyclic patterns ascribed as circadian rhythms. Through daily morphological and physiological adaptations, e.g., leaf angle and chloroplast movement, the plant is able to adapt to the fluctuating natural environment by predicting daily changes but also by anticipating regular natural events such as dawn (Dodd et al., 2015). In addition to circadian control, some processes respond cyclically to metabolic feedback; for example, in natural conditions, CO₂ assimilation usually follows a daily pattern characterized by an initial “photosynthesis activation” at dawn, a maximum CO₂ fixation at mid-morning, and a decline from midday (midday depression) (Koyama and Takemoto, 2014; Maai et al., 2019). After the midday depression, the photosynthetic activity declines until the dark when the nocturnal process of starch consumption is under circadian oscillator control (Haydon et al., 2013). In controlled environments lacking daylight, complete control of plant rhythms is possible and may be a route to further optimization of light treatments.

In this study, we used a dose-response curve to assess the relationship between irradiance of a broad-spectrum (PAR) LED array and the yield and pigmentation in Lollo rosso. From this curve, we identified a suitable treatment that produced a good combination of yield and morphology that was used to study the effects of PAR plus supplemental R and B on yield and pigmentation. We used supplemental B to examine the effects of duration and to identify a minimal active treatment for further diel studies. We hypothesized that the effectiveness of supplemental B would vary between different diel treatment periods.

MATERIALS AND METHODS

Plant Material and Growth Conditions

Four separate experiments were performed in the same controlled growth room with an 18-h and 6-h light and dark photoperiod. The walls were covered by white reflective sheets (ORCA grow film, California Grow Films LLC), and atmospheric conditions were monitored using Tinytag Ultra 2 (Gemini Data Loggers, Chichester, United Kingdom) and Rotronic CL11 (Rotronic Instruments Ltd., United Kingdom). The average air temperature was $21.9 \pm 0.6^\circ\text{C}$, relative humidity was $58.5 \pm 4.8\%$, and CO_2 was 470.5 ± 2.4 ppm; averaged environmental values for individual experiments are detailed in **Supplementary Table 1**.

Seeds of red lettuce Lollo rosso (Antoniet RZ seeds from Rijk Zwaan, De Lier, Netherlands) were sown in 155 g of sieved John Innes No. 3 soil-based compost. Water-holding “field” capacity of the compost was calculated following the gravimetric method for soil moisture determination (Reynolds, 1970). Pots were filled, saturated with water, covered with plastic film, and left to drain at room temperature ($20 \pm 5^\circ\text{C}$). After 24 h, pot weight was noted and pots were incubated in the oven at 105°C . Every 24 h, pots were weighed until stable dry weight was reached. The dry and wet weights were used to estimate the weight of pots, and soil at approximately 0% and 100% water holding capacity and capacities in between these extremes were estimated as a linear proportion of the difference between these values. Pots were individually irrigated to 80% water holding capacity (205 g) every 48 h until harvest at 30 days after sowing (DAS).

Light intensity and spectral composition of the treatments were measured using the spectroradiometer SpectraPen LM 500 (cosine-corrected, 380–780 nm; Photon Systems Instruments, Drásov, Czechia) (**Supplementary Figure 1**).

Broad-Spectrum LED Light Response Curve

Seeds were germinated and grown under two broad-spectrum (PAR) LED arrays (EP006, 380–760 nm, Shenzhen Herifi Technology Co., Ltd., China) (**Supplementary Figure 1A**) for 30 days. A total of twelve irradiance treatments (5, 10, 15, 30, 40, 60, 80, 100, 120, 140, 160, 180 W m^{-2} or 25, 30, 75, 150, 200, 300, 400, 500, 600, 700, 800, 900 $\mu\text{mol m}^{-2} \text{s}^{-1}$) were obtained from the same broad spectrum by adding different layers of muslin cloth as a neutral density filter between source and individual plants. Each treatment was replicated three times and the plants were used for FW determination.

Supplementing Broad-Spectrum (PAR) LED Arrays With Blue and Red LED Light

Seeds were germinated and grown for 15 days under a broad-spectrum PAR LED source (EP006, 380–760 nm, Shenzhen Herifi Technology Co., Ltd., China) with a photosynthetic photon flux density (PPFD) of $300 \mu\text{mol m}^{-2} \text{s}^{-1}$ (P60). After growth under the P60 array, groups of 9 randomly selected plants were moved under different spectral treatments comprising either double the PAR irradiance (P120), the same PAR irradiance supplemented with R LEDs (P60 + R), or the same PAR irradiance supplemented

with B LEDs (P60 + B). Supplemental B and R were provided from mixed arrays of two LEDs centered at 430 and 460 nm (B) and 630 and 660 nm (R) (**Supplementary Figures 1B,C**). Treatments P120, P60 + R, and P60 + B provided approximately the same PPFD ($500 \mu\text{mol m}^{-2} \text{s}^{-1}$). Plants were moved to one of the three supplemental treatments either at 15 DAS or 26 DAS, leaving the remainder under P60 radiation, thus plants were grown under supplemental treatments for either 15 days or 4 days.

Supplementing Broad-Spectrum (PAR) LEDs With Different Durations of B LEDs

Seeds were germinated and grown for 15 days under a broad-spectrum PAR LED source (EP006, 380–760 nm, Shenzhen Herifi Technology Co., Ltd., China) with a photosynthetic photon flux density (PPFD) of $300 \mu\text{mol m}^{-2} \text{s}^{-1}$ (P60). Plants were randomly selected to either remain under PAR treatment or were grown under similar PAR LEDs supplemented with B radiation to reach a PPFD of $500 \mu\text{mol m}^{-2} \text{s}^{-1}$ (total B accounted for 44% of the emission spectrum). Supplemental B LED treatments varied from a high daily light integral (DLI) B treatment in which plants were transferred to PAR plus supplemental B for the remaining 15 days of the experiment to a minimal DLI B treatment whereby plants remained under P60 for 29 days and received 1 h of PAR60 plus supplemental B LED treatment on the final day (see **Table 1** for full range of treatments).

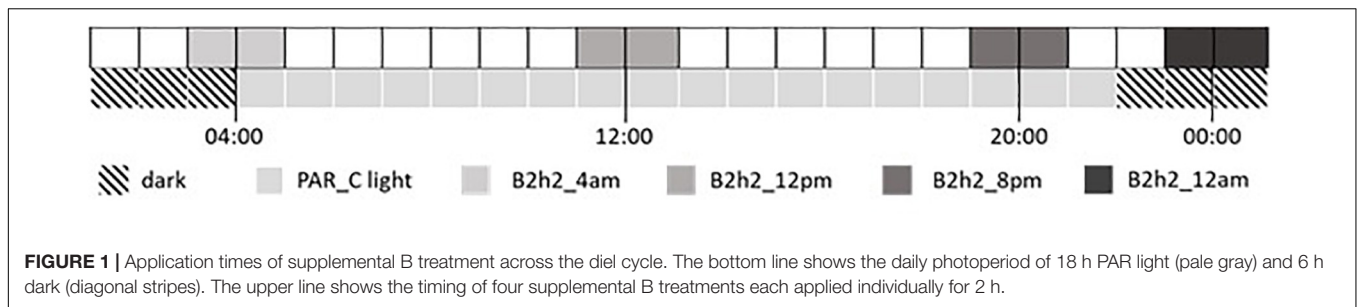
Interaction of Supplemental B LED Light and the Diel Cycle

Seeds were germinated and grown for 15 days under a broad-spectrum PAR LED source (EP006, 380–760 nm, Shenzhen Herifi Technology Co., Ltd., China) with a photosynthetic photon

TABLE 1 | Duration, description, and daily light integrals (DLI) of treatment regimes with PAR plus supplemental B applied for different periods across a 30-days growth period.

Treatments	PAR radiation	Supplemental B radiation	Total DLI ($\text{mol m}^{-2} \text{d}^{-1}$)	Total B DLI ($\text{mol m}^{-2} \text{d}^{-1}$)
P60	30 days (18 h photoperiod)	–	19.44	6.09
B15D	15 days (18 h photoperiod)	15 days (18 h photoperiod)	26.50	14.09
B4D	26 days (18 h photoperiod)	4 days (18 h photoperiod)	21.30	8.22
B2D	28 days (18 h photoperiod)	2 days (18 h photoperiod)	20.38	7.16
B1D	29 days (18 h photoperiod)	1 day (18 h photoperiod)	19.91	6.62
B9h	29 days + 9 h	9 h	19.68	6.36
B4h	29 days + 14 h	4 h	19.54	6.20
B2h	29 days + 16 h	2 h	19.49	6.15
B1h	29 days + 17 h	1 h	19.47	6.12

Daily light integral (DLI) was calculated by multiplying the instantaneous photon flux density (PPFD) ($\mu\text{mol m}^{-2} \text{s}^{-1}$) for the total time of the treatment application, then was divided by the number of growth days (30) in order to obtain the mol of photons per day reaching the plant.



flux density (PPFD) of $300 \mu\text{mol m}^{-2} \text{s}^{-1}$ (P60). Plants were randomly selected to either remain under PAR treatment or were grown for a further 15 days under similar PAR LEDs supplemented with B LED treatments (PPFD of $800 \mu\text{mol m}^{-2} \text{s}^{-1}$) for 2 h during different periods of the day–night cycle. Four treatment times were tested placing the 2-h B treatment at the beginning, middle, and end of the 18-h light period and in the middle of the 6-h dark period (Figure 1).

Sampling and Measurements of Plant Morphological and Physiological Parameters

Chlorophyll *a* fluorescence was assessed from leaf number four using a portable Handy PEA continuous excitation chlorophyll fluorimeter (Hansatech Instruments Ltd., King's Lynn, United Kingdom). First, light-adapted measurements were taken, then dark-adapted measurements after 30 min of dark adaptation using the manufacturer's leaf clips. Measurements were always in the morning just after 10:00, except when treatments required measurements at different specific times.

Rosette images were taken using a fixed focal length digital camera and stand. Images were used for rosette area determination using the "Shape descriptor" plugin in ImageJ software (version 1.52a) (Schneider et al., 2012). The rosette was harvested from just above the cotyledon node and immediately weighed for fresh weight (FW). The entire rosette was then placed in a paper bag and dried to constant weight at 60°C to determine dry weight (DW).

A random selection of plants not used for yield determination was harvested for biochemical analyses at the end of the experiment (day 30). Fully expanded leaves, developmentally the third and fourth leaves, were excised, the midrib was removed, and tissue was immediately frozen in liquid nitrogen before storage at -80°C until analyzed. Prior to analysis, samples were freeze-dried and cold milled to a fine powder in an automated sample grinder (Labman Automation Ltd., Middlesbrough, United Kingdom) for 90 s at -70°C .

Extraction and Quantification of Anthocyanin Content

Anthocyanins were quantified as a single peak cyanidin 3-malonylglucoside [reported as main anthocyanin in Lollo rosso (Ferrerres et al., 1997)] confirmed by fragmentation pattern and mass spectroscopy. Lyophilized powdered leaf material

(30 mg) was extracted by shaking in acidified methanol solution (methanol: water: acetic acid; 70: 28.5: 1.5) for 30 min at room temperature. The sample was then centrifuged for 10 min at $1500 \times g$, and the extract was collected and evaporated in a centrifugal evaporator (Jouan, RC 10.22). The concentrated extract was then purified by Solid Phase Extraction using sep-pak cartridges (500 mg Sep-Pak C_{18} 3 cc Vac RC cartridge, Waters Ltd., Elstree, United Kingdom). The final extract was analyzed by reversed-phase HPLC using a Waters system equipped with a 996 photodiode detector array (PDA) and a Nova-Pak C_{18} radial compression column ($8 \text{ mm} \times 100 \text{ mm}$, particle size $4 \mu\text{m}$; Waters Ltd., Elstree, United Kingdom). The column was equilibrated with 20% solvent A (5% acetic acid) at a flow rate of 2 ml min^{-1} . Compounds were eluted by linear gradient to 60% solvent B (100% methanol) over 20 min and monitored from 240 to 600 nm with the detection wavelength set to 525 nm. Anthocyanins were quantified from peak areas using an external standard curve using a cyanidin chloride standard (Sigma-Aldrich Company Ltd.) which had a very similar retention time.

Statistical Analysis

All the data were statistically analyzed using Microsoft Excel 2016 and R studio [R version 3.5.2 (2018-12-20), "Eggshell Igloo"] with packages agricolae, car, ggplot2, and segmented (Muggeo, 2003; De Mendiburu, 2020; Fox et al., 2020; Wickham et al., 2020). For the measured parameters, data were analyzed by one-way ANOVA and the means were compared by least significance difference (LSD), at 5% significance level.

RESULTS

Yield and Morphological Responses to Varying Irradiance of Broad-Spectrum (PAR) LEDs

Using a broad-spectrum (PAR) LED array, to provide a range of irradiances between 5 and 180 W m^{-2} , demonstrated a highly plastic response in the pigmented lettuce Lollo rosso. Rosette fresh weight (FW) increased logarithmically with increasing irradiance. At lower irradiances, between 5 and 40 W m^{-2} , rosette FW increased linearly with increasing irradiance at an average rate of $0.4 \text{ g FW per W m}^{-2}$. Higher irradiances, between 40 and 80 W m^{-2} , produced $0.1 \text{ g per W m}^{-2}$, while irradiances

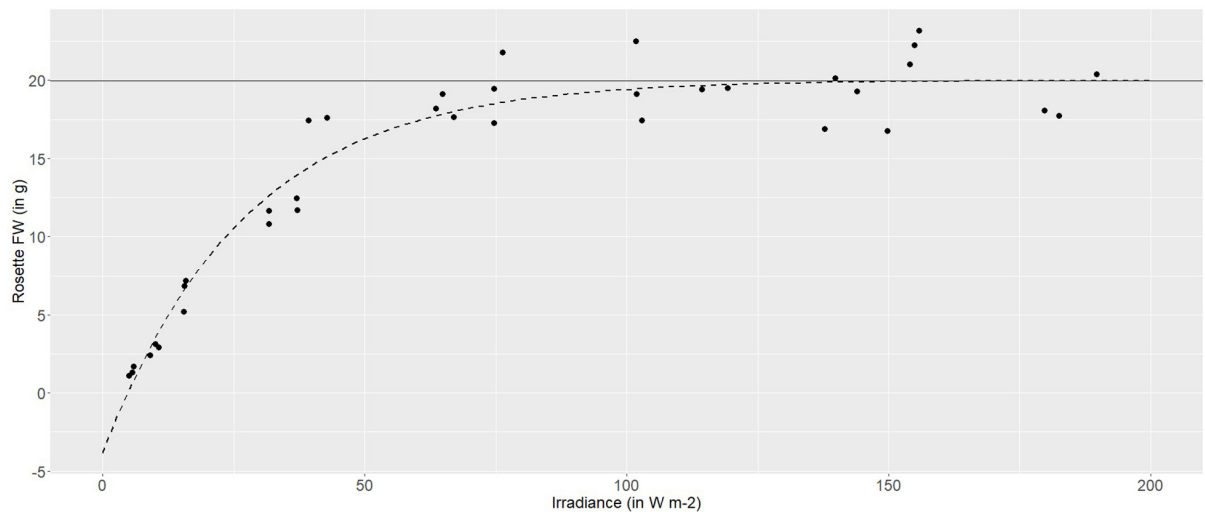


FIGURE 2 | Non-linear (logarithmic) asymptotic regression between the fresh weight (g) of Lollo rosso and irradiance (W m^{-2}). In dark gray is the horizontal asymptote, and in dashed dark gray is the model-based regression curve.

TABLE 2 | Morphological responses of Lollo rosso to varying the incident irradiance of a broad-spectrum (PAR) LED array.

Averaged light treatments (W m^{-2})	Averaged light treatment ($\mu\text{ mol m}^{-2} \text{ s}^{-1}$)	Leaf number***	Rosette area*** (cm^2)	Anthocyanin*** ($\text{mgCYA g}^{-1} \text{ DW}$)
5.5	27.0	5.0 ± 0.0^f	73.1 ± 12.7^f	0.19 ± 0.01^f
9.9	48.2	6.3 ± 0.3^{ef}	131.2 ± 14.0^e	0.28 ± 0.01^f
15.6	75.4	6.7 ± 0.3^{def}	211.7 ± 4.5^a	0.18 ± 0.02^f
33.5	161.6	7.7 ± 0.3^{de}	171.3 ± 7.1^{cd}	2.07 ± 0.14^e
39.7	186.9	8.7 ± 0.3^{cd}	200.1 ± 6.2^{abc}	4.21 ± 0.13^d
65.1	329.7	10.7 ± 1.2^{bc}	203.4 ± 5.5^{ab}	5.46 ± 0.08^c
75.3	387.1	13.0 ± 0.6^a	186.6 ± 9.3^{abcd}	5.72 ± 0.09^{bc}
102.2	387.1	12.7 ± 0.9^{ab}	183.1 ± 10.7^{abcd}	5.75 ± 0.09^{bc}
125.9	517.9	12.7 ± 1.5^{ab}	194.9 ± 16.8^{abc}	6.03 ± 0.53^{abc}
142.4	731.1	11.7 ± 0.3^{ab}	157.2 ± 5.8^{de}	6.55 ± 0.22^{ab}
154.9	800.0	13.0 ± 0.6^a	180.8 ± 6.6^{bcd}	6.87 ± 0.63^a
184.0	942.4	12.7 ± 0.3^{ab}	141.8 ± 11.1^e	6.21 ± 0.74^{abc}

Values are reported as mean \pm standard error of the mean. Different letters within columns indicate significant treatment differences at $P < 0.05$, as determined by analysis of variance (ANOVA) and Fisher's least significant difference (LSD) test. Significance codes: 0.000 "****"; $n = 3$.

of 100–120 W m^{-2} produced a maximum rosette FW of 20 g and growth was asymptotic at higher irradiances (Figure 2).

Increasing irradiance induced significant changes to morphological responses such as rosette area ($P = 4.7 \times 10^{-8}$), leaf number ($P = 2.9 \times 10^{-9}$), and leaf pigmentation ($P = 4.5 \times 10^{-5}$). Rosette area almost tripled from 5 ($73.1 \pm 12.7 \text{ cm}^2$) to 15 W m^{-2} ($211.7 \pm 4.5 \text{ cm}^2$), and plants growing under the lower irradiances displayed a prostrate leaf morphology. Rosette area decreased linearly between light energy of 60 and 180 W m^{-2} to a rosette area of $141.8 \pm 11.1 \text{ cm}^2$ (Table 2). Anthocyanins were difficult to detect in plants grown under very low irradiance (up to 15 W m^{-2} , 0.18–0.28 $\text{mg g}^{-1} \text{ DW}$). Light levels of 30 W m^{-2} produced increased accumulation of anthocyanin content ($2.07 \pm 0.14 \text{ mg g}^{-1} \text{ DW}$) and reached a maximum under 160 W m^{-2} ($6.87 \pm 0.63 \text{ mg g}^{-1} \text{ DW}$) (Table 2).

Supplementing Broad-Spectrum (PAR) LED Arrays With Blue and Red LED Light

There was no significant effect on rosette FW and DW of supplementing broad-spectrum (PAR) LED arrays with R and B LEDs at the irradiances tested (Table 3), but morphology was affected especially rosette area and leaf pigmentation. Rosette area was reduced by the longer (15 days) supplemental treatments ($P = 0.001$). A 15-days supplementation with B LEDs produced the most compact rosette with the lowest area ($167.39 \pm 3.13 \text{ cm}^2$), next lowest was 15-days supplementation with PAR LEDs (188.62 ± 6.09), and 15-days supplementation with R LEDs produced a larger rosette area (193.43 ± 4.63). The largest rosette area was produced by Lollo rosso growing under the 4-days supplementation with R LEDs. Accumulation of anthocyanins was enhanced under supplemental B only and was greater in leaves exposed to 4 days than 15 days of supplemental

B LEDs (13.00 ± 0.44 and 9.60 ± 0.65 mg g⁻¹ DW after 4 days and 15-days B supplementation, respectively) (Table 3) ($P = 2.2 \times 10^{-16}$).

Supplementing Broad-Spectrum (PAR) LEDs With Different Durations of B LEDs

The duration of supplemental B radiation had a significant effect on rosette FW ($P = 1.9 \times 10^{-14}$) and DW ($P = 7.2 \times 10^{-8}$) of Lollo rosso (Table 4). Longer durations of supplemental B increased biomass accumulation which was greatest and asymptotic under 1, 2, 4, and 15 days of supplemental B (19.1 to 20.4 g). Rosette DW followed a similar trend to FW, but the greatest DW was measured after 1 day of supplemental B. Rosette area in general decreased with increasing duration of supplemental B. The smallest rosette areas (1.8-fold smaller

than rosettes from control plants) were measured in plants treated with the longest duration of supplemental B (15 days, 146.8 ± 8.5 cm²), and the largest rosette area was measured in plants treated with 1 day of supplemental B (181.3 ± 11.5 cm²); rosette areas from all the intermediate treatments did not significantly differ (Table 4).

Supplemental B did not have a significant effect on chlorophyll *a* fluorescence and the maximum quantum efficiency of PSII photochemistry in the dark (F_V/F_M) approximated to a similar value of 0.83 in all measured plants. The amount of light energy dissipated via non-photochemical quenching generally increased with duration of supplemental B, but the differences were not statistically significant.

All supplemental B treatments greater than 1-h duration resulted in significant increases in anthocyanin content compared to levels in leaves from control plants lacking supplemental B ($P = 3.6 \times 10^{-14}$). The anthocyanin content increased approximately linearly with increasing durations of supplemental B from 2 h (27%) to 4 days (224%) (Table 4). Anthocyanin accumulation declined slightly in leaves exposed to the longest, 15-days, supplemental B treatment (187%). The amount of anthocyanin in leaves exposed to the shortest supplemental B treatment (1 h) did not statistically differ from that in control leaves.

TABLE 3 | Fresh and dry weight yield, rosette area, and anthocyanin content of leaf tissue of Lollo rosso lettuce growing under $300 \mu\text{mol m}^{-2} \text{s}^{-1}$ broad-spectrum (PAR) LED (P60) for 15 days and followed by a further 15 days under P60 or P60 plus supplemental PAR, Red (R) or Blue (B) LEDs for either 15 days or 4 days.

Supplemental treatments	Rosette area** (cm ²) (n = 3)	Fresh weight (g) (n = 9)	Dry weight (g) (n = 6)	Anthocyanin*** (mgCYA g ⁻¹ DW) (n = 3)
–	239.9 ± 14.4 ^a	20.4 ± 1.0	1.46 ± 0.09	4.96 ± 0.20 ^c
15D P60	188.6 ± 6.1 ^{bc}	17.9 ± 1.2	1.26 ± 0.10	4.14 ± 0.16 ^{cd}
4D P60	240.6 ± 11.3 ^a	19.9 ± 0.7	1.47 ± 0.05	4.42 ± 0.15 ^{cd}
15D R	193.4 ± 4.6 ^{bc}	21.7 ± 1.1	1.50 ± 0.18	3.77 ± 0.40 ^d
4D R	246.3 ± 21.4 ^a	19.8 ± 1.6	1.45 ± 0.09	3.75 ± 0.49 ^d
15D B	167.4 ± 3.1 ^c	20.3 ± 0.5	1.44 ± 0.04	9.60 ± 0.65 ^b
4D B	221.1 ± 7.0 ^{ab}	19.8 ± 0.7	1.67 ± 0.08	13.00 ± 0.44 ^a

Values are reported as mean ± standard error of the mean. Different letters within columns indicate significant treatment differences at $P < 0.05$, as determined by analysis of variance (ANOVA) and Fisher's least significant difference (LSD) test. Significance codes: 0.000 "****", 0.001 "****".

TABLE 4 | Fresh and dry weight yield, rosette area, and anthocyanin content of leaf tissue of Lollo rosso lettuce growing under $300 \mu\text{mol m}^{-2} \text{s}^{-1}$ broad-spectrum (PAR) LED (P60) for 15 days and then a further 15 days under P60 with different durations of supplemental Blue (B) LEDs.

Treatments	Rosette area* (cm ²) (N = 3)	Fresh weight*** (g) (N = 8)	Dry weight*** (g) (N = 4)	Anthocyanin*** (mgCYA g ⁻¹ DW) (N = 3)
P60	202.9 ± 19.4 ^a	11.8 ± 0.5 ^c	0.85 ± 0.13 ^d	3.98 ± 0.21 ^g
B15D	146.8 ± 8.5 ^c	20.4 ± 0.5 ^a	1.46 ± 0.07 ^{bc}	11.44 ± 0.39 ^b
B4D	174.9 ± 1.5 ^b	20.2 ± 0.7 ^a	1.60 ± 0.08 ^{abc}	12.91 ± 0.43 ^a
B2D	169.7 ± 5.1 ^{bc}	20.3 ± 1.3 ^a	1.72 ± 0.10 ^{ab}	8.48 ± 0.48 ^c
B1D	181.3 ± 11.5 ^{ab}	19.1 ± 1.6 ^a	1.76 ± 0.21 ^a	7.53 ± 0.29 ^d
B9h	166.2 ± 2.7 ^{bc}	15.4 ± 0.8 ^b	1.44 ± 0.06 ^{bc}	7.09 ± 0.09 ^{de}
B4h	168.8 ± 4.3 ^{bc}	15.7 ± 0.7 ^b	1.35 ± 0.06 ^c	6.44 ± 0.05 ^e
B2h	161.4 ± 3.7 ^{bc}	11.2 ± 0.4 ^c	0.79 ± 0.03 ^d	5.04 ± 0.15 ^f
B1h	170.0 ± 4.0 ^{bc}	12.0 ± 0.6 ^c	0.85 ± 0.06 ^d	3.69 ± 0.04 ^g

Values are reported as mean ± standard error of the mean. Different letters within columns indicate significant treatment differences at $P < 0.05$, as determined by analysis of variance (ANOVA) and Fisher's least significant difference (LSD) test. Significance codes: 0.000 "****", 0.01 "****".

Interaction of Supplemental B LED Light and the Diel Cycle

When Lollo rosso was grown in an 18-h light, 6-h dark cycle, the timing of a 2-h supplemental B treatment within the light and dark periods had a significant effect on rosette FW ($P = 0.008$). When supplemental B was applied in the middle of the dark period (12 am), rosette FW was greatest. When supplemental B was applied at the end of the light cycle (8 pm), rosette FW was lowest and did not significantly differ from control plants lacking supplemental B. Similar trends were seen in rosette DW ($P = 0.05$) (Table 5).

Anthocyanin content was significantly higher when supplemental B was supplied in the middle of the dark period (9.81 ± 0.37 mg g⁻¹ DW) than all other treatment and control plants ($P = 3.7 \times 10^{-8}$) (Table 5). Supplementing with 2-h B at the beginning (4 am) and middle (12 pm) of the light cycle (6.60 ± 0.29 mg g⁻¹ DW and 7.50 ± 0.48 mg g⁻¹ DW, respectively) resulted in significantly higher anthocyanin accumulation than control plants (4.00 ± 0.14 mg g⁻¹ DW), but anthocyanin contents of control plants and plants treated with supplemental B at the end of the light cycle were not significantly different.

F_V/F_M was similar between differently treated plants and approximated the optimal value. Maximum operating efficiency of PSII photochemistry in the light (F_V/F_M') was lowest under midday supplementation and highest under midnight supplementation ($P = 0.010$). Performance index (PI) was similar across the diverse treatments, except the significantly lower values measured when supplementation was applied at the end of the light cycle ($P \geq 0.000$). Non-photochemical quenching (NPQ) was low and similar in leaves grown in control (P60) and

TABLE 5 | Fresh and dry weight yield, and anthocyanin content of leaf tissue of Lollo rosso lettuce growing under 300 $\mu\text{mol m}^{-2} \text{s}^{-1}$ broad-spectrum (PAR) LED (P60) for 15 days and then a further 15 days under P60 and P60 supplemented with Blue (B) LEDs for 2 h at four different periods.

Treatments	Fresh weight** (g) (n = 8)	Dry weight- (g) (n = 4)	Anthocyanin*** (mgCYA g ⁻¹ DW) (n = 4)
P60	10.6 ± 0.6 ^b	0.79 ± 0.09 ^{ab}	4.00 ± 0.14 ^c
P60 + 2hB (12 am)	12.4 ± 0.4 ^a	0.92 ± 0.04 ^a	9.81 ± 0.32 ^a
P60 + 2hB (4 am)	10.1 ± 0.7 ^b	0.73 ± 0.06 ^b	6.60 ± 0.29 ^b
P60 + 2hB (12 pm)	11.0 ± 0.4 ^{ab}	0.75 ± 0.04 ^{ab}	7.50 ± 0.48 ^b
P60 + 2hB (8 pm)	9.6 ± 0.6 ^b	0.71 ± 0.03 ^b	4.43 ± 0.44 ^c

Values are reported as mean ± standard error of the mean. Different letters within columns indicate significant treatment differences at $P < 0.05$, as determined by analysis of variance (ANOVA) and Fisher's least significant difference (LSD) test. Significance codes: 0.000 "****," 0.001 "****," and 0.05 ".,".

TABLE 6 | Chlorophyll fluorescence parameters of leaf tissue of Lollo rosso lettuce growing under 300 $\mu\text{mol m}^{-2} \text{s}^{-1}$ broad-spectrum (PAR) LED (P60) for 15 days and then a further 15 days under P60 and P60 supplemented with Blue (B) LEDs for 2 h at four different periods.

Treatments	F _V /F _M **	F _V /F _M	PI**	NPQ*
P60	0.77 ± 0.01 ^{ab}	0.85 ± 0.01	3.01 ± 0.30 ^a	0.28 ± 0.02 ^{bc}
P60 + 2h B (12 am)	0.77 ± 0.01 ^a	0.85 ± 0.00	3.70 ± 0.22 ^a	0.23 ± 0.05 ^c
P60 + 2h B (4 am)	0.75 ± 0.01 ^{bc}	0.84 ± 0.00	3.08 ± 0.43 ^a	0.40 ± 0.04 ^{ab}
P60 + 2h B (12 pm)	0.74 ± 0.01 ^c	0.84 ± 0.01	3.77 ± 0.38 ^a	0.50 ± 0.06 ^a
P60 + 2h B (8 pm)	0.74 ± 0.02 ^{bc}	0.84 ± 0.01	1.74 ± 0.11 ^b	0.43 ± 0.08 ^{ab}

Values are reported as mean ± standard error of the mean. Different letters within columns indicate significant treatment differences at $P < 0.05$, as determined by analysis of variance (ANOVA) and Fisher's least significant difference (LSD) test. Significance codes: 0.001 "****," 0.01 "****," n = 4.

midnight supplemental B treatments but was significantly higher in all three treatments when B was supplemented during the light cycle ($P = 0.015$) (Table 6).

DISCUSSION

The use of LED lighting is revolutionizing the provision of light to controlled environment agriculture (Pattison et al., 2018). Light intensity has a significant effect on plant growth and morphology (Poorter et al., 2019), and to fully exploit the flexibility afforded by new LED-based lighting systems will involve investigation of the interactions between crop production and variation in irradiance and spectrum (Gómez and Izzo, 2018). Less is known about the potential for further optimization through interactions between light and other factors such as temperature (Franklin et al., 2014) and circadian rhythms (Belbin et al., 2017).

Here we used Lollo rosso, a fast-growing commercial lettuce variety, as a model system to study how irradiance, spectrum, duration, and rhythm of LED light may be used to affect crop growth. We studied aspects of both yield and quality, the

latter through the accumulation of anthocyanin pigment. By growing Lollo rosso under a range of PAR irradiance levels, limiting, optimal, and saturating light levels were identified. Light intensity of over 100 W m^{-2} (PAR $\sim 520 \mu\text{mol m}^{-2} \text{s}^{-1}$) produced asymptotic growth, suggesting that photosynthesis and or partitioning of photosynthate into harvestable yield is saturated at such high irradiances. Light exceeding the limiting light levels is more likely to activate photoprotective responses (Robinson, 2001), and light not directly converted to harvestable yield may impact crop quality through alterations in morphology and composition. Rosette area, for instance, decreased in response to irradiance. Under low irradiance light conditions (5–15 W m^{-2}), a pale loose-leaf head developed which lacked measurable levels of anthocyanins; such phenotypes are associated with shade conditions to optimize light capture (Björkman and Demmig-Adams, 1995). A more compact rosette head formed and anthocyanin content doubled at moderate light levels (30–40 W m^{-2}). At higher irradiances, the rosette area decreased, resulting in more compact rosettes, presumably as a result of hypocotyl length and leaf angle reduction, both traits reported to be protective strategies that decrease incident light interception (Rama Das, 2006; Arsovski et al., 2012). Combining assessments of yield and morphology we concluded that PAR LEDs at 60 W m^{-2} (approximately 300 $\mu\text{mol m}^{-2} \text{s}^{-1}$) produced a good combination of yield and morphology in Lollo rosso, and this treatment was chosen to examine supplementing broad spectrum LED light with additional narrow-band LEDs.

Maximal leaf absorbance occurs in the B and R wavelengths due to the absorption peaks of chlorophylls a and b (428–453 and 642–661 nm) and carotenoids (400–500 nm) (McCree, 1971; Barber, 2009). Hence, the previously identified PAR LED irradiance level was supplemented with R or B radiation to reach a higher, but not saturating, light intensity (approximately 500 $\mu\text{mol m}^{-2} \text{s}^{-1}$). Our results demonstrated that supplementation of a PAR LED array with additional R or B LEDs had no significant effect on Lollo rosso biomass accumulation. However, supplemental B was effective at stimulating other desirable plant traits including rosette area and anthocyanin content. The reduced rosette area under supplemental B probably resulted from previously demonstrated effects of B inhibiting stem elongation and controlling leaf orientation (Huché-thélier et al., 2016). The increase in cyanidin content compared to PAR treatments lacking supplemental B was greater in the shorter B treatment. In contrast, NPQ increased with duration of supplemental B and a similar contrast was reported in young leaves of *Acmena acuminatissima* in contrasting seasons by Zhu et al. (2018). The phenotype of decreasing pigmentation and increasing stress may be due to alternative acclimation responses (Nogués et al., 1998), with the plant investing more in internal and constitutive protective mechanisms such as energy dissipation as heat through the xanthophyll cycle or rearrangements of photosystem machinery, rather than adopting largely preventive strategies through increased pigmentation (Steyn et al., 2002; Bailey et al., 2004; Li et al., 2009).

Having established the potential for shorter-duration supplemental B to improve pigmentation and morphology,

we next determined the relationship between duration of supplemental B treatment and pigmentation to identify a minimal treatment condition. Identifying a short enough treatment would enable us to further investigate the interaction of supplementation and the diel growth cycle. If absorbed by plants, the highly energetic photons of short wavelength light may increase photoprotection responses and negatively impact plant growth (Landi et al., 2015). UV radiation, for example, has been widely studied in this respect and stimulates the accumulation of secondary metabolites in plants (Schreiner et al., 2012) and inhibits growth in many plant species including lettuce (Tsormpatsidis et al., 2010; Lee et al., 2013). In contrast to UV, B wavelengths are photosynthetically active (Zhu et al., 2018), B LEDs are readily available commercially and can produce light at a higher efficiency (93%) compared with 81% for R LEDs (Kusuma et al., 2020). Thus, supplementation with B LEDs represents a suitable system to examine ways to increase photoassimilation and increase pigmentation while minimizing the negative impacts of phototoxicity, non-photochemical quenching, and consequent reduction in yield.

Exposing Lollo rosso to supplemental B for decreasing periods of time (from 15 days to 1 h) produced a distinct dose–response effect on the measured variables. Leaf anthocyanin content increased linearly from 2 h to 4 days of B supplementation. Rosette area and NPQ increased proportionally with treatment duration. The plateau in yield at around 1 day of supplemental B treatment suggests that the extra B photons were no longer contributing to net biomass accumulation at longer durations and the decrease in leaf anthocyanin content after the longest B treatment may be caused by acclimation of the plant to the high light environment. A similar response was reported by Taulavuoria et al. (2016) in which many compounds in basil decreased after B treatment for 48 days compared to a shorter 36-days treatment.

A minimal treatment of 2-h supplemental B was sufficient to significantly increase cyanidin content but had no significant effect on biomass. Thus, in the next test, the 2-h B treatment was adjusted to deliver an equivalent DLI to the most effective 4 days treatment ($\sim 21.3 \text{ mol m}^{-2} \text{ d}^{-1}$, **Table 4**) and broad-spectrum PAR LEDs were supplemented with the resulting 2-h B at different times in the diel cycle. Increasing daily light integral (DLI), or the total sum of radiation in a 24-h period, allows higher radiation sums with lower PPFD, thus avoiding negative effects of saturating light levels and increasing lettuce fresh and dry mass (Kelly et al., 2020). Supplemental B at midday resulted in the highest NPQ which could be associated with midday depression of photosynthesis, stomatal closure, and reduced ability to dissipate light energy photochemically (Koyama and Takemoto, 2014). The lower total PPFD in the night break treatment (545 compared to $830 \mu\text{mol m}^{-2} \text{ s}^{-1}$) may allow most incident B photons to be photochemically quenched explaining the high and unaffected F_V/F_M' and the low level of NPQ. B light application in the night period was reported to increase carbon export and enhance fruit production (Lanoue et al., 2019), and the night break produced the highest biomass of our diel treatments. The higher percentage of B in the total PPF, 100% in the midnight treatment compared to 75% in PAR

LEDs supplemented with B radiation, may explain the greater effectiveness at inducing anthocyanins of the supplemental B night break since anthocyanin content has been reported to increase with the percentage of B (Hernández et al., 2016).

Our data demonstrates the effectivity of supplemental B LEDs in stimulating leaf cyanidin accumulation while enhancing plant growth in Lollo rosso. Our results suggest that application of short-duration supplemental B LED light to PAR arrays has beneficial effects on Lollo rosso yield, morphology, and pigmentation and further that shorter duration supplementation may be made even more effective if applied during night breaks. Such studies will help producers improve crop quality and maximize returns on energy input from supplemental LED lighting utilizing visible wavelengths.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/**Supplementary Material**, further inquiries can be directed to the corresponding author.

AUTHOR CONTRIBUTIONS

LC: conceptualization, methodology, investigation, formal analysis, writing – original draft, and writing – reviewing and editing. ID: funding acquisition and supervision. PR: funding acquisition, conceptualization, resources, writing – original draft, writing – reviewing and editing, and supervision. All authors contributed to the article and approved the submitted version.

FUNDING

LC was funded by an Aberystwyth University studentship. ID and PR by the BBSRC Core Strategic Programme in Resilient Crops BBS/E/W/0012843A.

ACKNOWLEDGMENTS

We thank Dr. Ana Winters and Dr. Barbara Hauck (Aberystwyth University) for their assistance in anthocyanin extraction and analysis.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fpls.2020.598082/full#supplementary-material>

Supplementary Figure 1 | Light spectral composition of the PAR array (**A**, solid line in pink), the blue LEDs (**B**, dashed line in blue), and the red LEDs (**C**, dotted

line in red). Each line represents the average of three measurements recorded at different spots of the plant canopy.

Supplementary Table 1 | Environmental data for the experiments reported. Values are reported as mean \pm standard error of the mean.

REFERENCES

- Adams, W. W., and Demmig-Adams, B. (1992). Operation of the xanthophyll cycle in higher plants in response to diurnal changes in incident sunlight. *Planta* 186, 390–398. doi: 10.1007/BF00195320
- Arsovski, A. A., Galstyan, A., Guseman, J. M., and Nemhauser, J. L. (2012). Photomorphogenesis. *Am. Soc. Plant Biol.* 150, 202–212. doi: 10.1199/tab.0147
- Bailey, S., Horton, P., and Walters, R. G. (2004). Acclimation of *Arabidopsis thaliana* to the light environment: the relationship between photosynthetic function and chloroplast composition. *Planta* 218, 793–802. doi: 10.1007/s00425-003-1158-5
- Bantis, F., Smirnakou, S., Ouzounis, T., and Koukounaras, A. (2018). Current status and recent achievements in the field of horticulture with the use of light-emitting diodes (LEDs). *Sci. Horticult.* 235, 437–451. doi: 10.1016/j.scienta.2018.02.058
- Barber, J. (2009). Photosynthetic energy conversion: natural and artificial. *Chem. Soc. Rev.* 38, 185–196. doi: 10.1039/b802262n
- Belbin, F. E., Noordally, Z. B., Wetherill, S. J., Atkins, K. A., Franklin, K. A., and Dodd, A. N. (2017). Integration of light and circadian signals that regulate chloroplast transcription by a nuclear-encoded sigma factor. *New Phytol.* 213, 727–738. doi: 10.1111/nph.14176
- Benke, K., and Tomkins, B. (2017). Future food-production systems: vertical farming and controlled-environment agriculture. *Sustainability* 13, 13–26. doi: 10.1080/15487733.2017.1394054
- Björkman, O., and Demmig-Adams, B. (1995). “Regulation of photosynthetic light energy capture, conversion, and dissipation in leaves of higher plants,” in *BT – Ecophysiology of Photosynthesis*, Vol. 100, eds E.-D. Schulze and M. M. Caldwell (Berlin: Springer). doi: 10.1007/978-3-642-79354-7_2
- Chen, X., Xue, X., Guo, W., Wang, L., and Qiao, X. (2016). Growth and nutritional properties of lettuce affected by mixed irradiation of white and supplemental light provided by light-emitting diode. *Sci. Horticult.* 200, 111–118. doi: 10.1016/j.scienta.2016.01.007
- Clavijo-Herrera, J., Van Santen, E., and Gómez, C. (2018). Growth, water-use efficiency, stomatal conductance, and nitrogen uptake of two lettuce cultivars grown under different percentages of blue and red light. *Horticulturae* 4:16. doi: 10.3390/horticulturae4030016
- De Mendiburu, F. (2020). *Package ‘agricolae’*. Available online at: <https://CRAN.R-project.org/package=agricolae> (accessed June 7, 2020).
- Dodd, A. N., Belbin, F. E., Frank, A., and Webb, A. A. R. (2015). Interactions between circadian clocks and photosynthesis for the temporal and spatial coordination of metabolism. *Front. Plant Sci.* 6:245. doi: 10.3389/fpls.2015.00245
- Ferreres, F., Gil, M. I., Castañer, M., and Tomás-Barberán, F. A. (1997). Phenolic metabolites in red pigmented lettuce (*Lactuca sativa*). Changes with minimal processing and cold storage. *J. Agricult. Food Chem.* 45, 4249–4254. doi: 10.1021/jf970399j
- Fox, J., Weisberg, S., Price, B., Adler, D., Bates, D., Baud-bovy, G., et al. (2020). *Package ‘Car’*. Available online at: <https://CRAN.R-project.org/package=car> (accessed September 29, 2020).
- Franklin, K. A., Toledo-Ortiz, G., Pyott, D. E., and Halliday, K. J. (2014). Interaction of light and temperature signalling. *J. Exp. Bot.* 65, 2859–2871. doi: 10.1093/jxb/eru059
- Fu, W., Li, P., and Wu, Y. (2012). Effects of different light intensities on chlorophyll fluorescence characteristics and yield in lettuce. *Sci. Horticult.* 135, 45–51. doi: 10.1016/j.scienta.2011.12.004
- Gómez, C., and Izzo, L. G. (2018). Increasing efficiency of crop production with LEDs. *AIMS Agricult. Food* 3, 135–153. doi: 10.3934/agrfood.2018.2.135
- Haydon, M. J., Hearn, T. J., Bell, L. J., Hannah, M. A., and Webb, A. A. R. (2013). Metabolic regulation of circadian clocks. *Semin. Cell Dev. Biol.* 24, 414–421. doi: 10.1016/j.semcdb.2013.03.007
- Hee, P., and Beom, L. (2001). Effects of CO₂ concentration, light intensity and nutrient level on growth of leaf lettuce in a plant factory. *Acta Horticult.* 548, 377–383.
- Hernández, R., Eguchi, T., Deveci, M., and Kubota, C. (2016). Tomato seedling physiological responses under different percentages of blue and red photon flux ratios using LEDs and cool white fluorescent lamps. *Sci. Horticult.* 213, 270–280. doi: 10.1016/j.scienta.2016.11.005
- Herron, A. H. A., and Mauzerall, D. (1972). The development of photosynthesis in a greening mutant of chlorella and an analysis of the light saturation curve. *Plant Physiol.* 50, 141–148.
- Huché-théliér, L., Crespel, L., Le, J., Morel, P., Sakr, S., and Leduc, N. (2016). Light signaling and plant responses to blue and UV radiations — Perspectives for applications in horticulture. *Environ. Exp. Bot.* 121, 22–38. doi: 10.1016/j.envexpbot.2015.06.009
- Ilieva, I., Ivanova, T., Naydenov, Y., Dandolov, I., and Stefanov, D. (2010). Plant experiments with light-emitting diode module in Svet space greenhouse. *Adv. Space Res.* 46, 840–845. doi: 10.1016/j.asr.2010.05.009
- Izzo, L. G., Arena, C., De Micco, V., Capozzi, F., and Aronne, G. (2019). Light quality shapes morpho-functional traits and pigment content of green and red leaf cultivars of *Atriplex hortensis*. *Sci. Horticult.* 246, 942–950. doi: 10.1016/j.scienta.2018.11.076
- Johkan, M., Shoji, K., Goto, F., Hahida, S., and Yoshihara, T. (2012). Effect of green light wavelength and intensity on photomorphogenesis and photosynthesis in *Lactuca sativa*. *Environ. Exp. Bot.* 75, 128–133. doi: 10.1016/j.envexpbot.2011.08.010
- Kaiser, E., Ouzounis, T., Giday, H., Schipper, R., Heuvelink, E., and Marcelis, L. F. M. (2019). Adding blue to red supplemental light increases biomass and yield of greenhouse-grown tomatoes, but only to an optimum. *Front. Plant Sci.* 9:2002. doi: 10.3389/fpls.2018.02002
- Kang, J. H., Krishna Kumar, S., and Atulba, S. (2013). Light Intensity and photoperiod influence the growth and development of hydroponically grown leaf lettuce in a closed-type plant factory system. *Horticult. Environ. Biotechnol.* 54, 501–509. doi: 10.1007/s13580-013-0109-8
- Kelly, N., Choe, D., Meng, Q., and Runkle, E. S. (2020). Promotion of lettuce growth under an increasing daily light integral depends on the combination of the photosynthetic photon flux density and photoperiod. *Sci. Horticult.* 272:109565. doi: 10.1016/j.scienta.2020.109565
- Kim, S. J., Hahn, E. J., Heo, J. W., and Paek, K. (2004). Effects of LEDs on net photosynthetic rate, growth and leaf stomata of chrysanthemum plantlets in vitro. *Sci. Horticult.* 101, 143–151. doi: 10.1016/j.scienta.2003.10.003
- Koyama, K., and Takemoto, S. (2014). Morning reduction of photosynthetic capacity before midday depression. *Sci. Rep.* 4, 1–6. doi: 10.1038/srep04389
- Kozai, T. (2013). Resource use efficiency of closed plant production system with artificial light: concept, estimation and application to plant factory. *Proc. Jpn. Acad. Ser. B Phys. Biol. Sci.* 89, 447–461. doi: 10.2183/pjab.89.447
- Kusuma, P., Pattison, P. M., and Bugbee, B. (2020). From physics to fixtures to food: current and potential LED efficacy. *Horticult. Res.* 7:56. doi: 10.1038/s41438-020-0283-7
- Landi, M., Tattini, M., and Gould, K. S. (2015). Multiple functional roles of anthocyanins in plant-environment interactions. *Environ. Exp. Bot.* 119, 4–17. doi: 10.1016/j.envexpbot.2015.05.012
- Lanoue, J., Zheng, J., Little, C., Thibodeau, A., Grodzinski, B., and Hao, X. (2019). Alternating red and blue light-emitting diodes allows for injury-free tomato production with continuous lighting. *Front. Plant Sci.* 10:1114. doi: 10.3389/fpls.2019.01114
- Lee, S., Seo, J. M., Lee, M. K., Chun, J. H., Antonisamy, P., Arasu, M. V., et al. (2013). Influence of different LED lamps on the production of phenolic compounds in common and Tartary buckwheat sprouts. *Indust. Crops Product.* 54, 320–326. doi: 10.1016/j.indcrop.2014.01.024

- Li, Q., and Kubota, C. (2009). Effects of supplemental light quality on growth and phytochemicals of baby leaf lettuce. *Environ. Exp. Bot.* 67, 59–64. doi: 10.1016/j.envexpbot.2009.06.011
- Li, Z., Wakao, S., Fischer, B. B., and Niyogi, K. N. (2009). Sensing and responding to excess light. *Ann. Rev. Plant Biol.* 60, 239–260. doi: 10.1146/annurev.arplant.58.032806.103844
- Lichtenthaler, H. K., Buschmann, C., and Rahmsdorf, U. (1980). "The Importance of blue light for the development of sun-type chloroplasts," in *The Blue Light Syndrome. Proceedings in Life Sciences*, ed. H. Senger (Berlin: Springer).
- Maai, E., Nishimura, K., Takisawa, R., and Nakazaki, T. (2019). Light stress-induced chloroplast movement and midday depression of photosynthesis in sorghum leaves. *Plant Product. Sci.* 00, 1–10. doi: 10.1080/1343943X.2019.1673666
- Matsuda, R., Ohashi-Kaneko, K., Fujiwara, K., Goto, E., and Kurata, K. (2004). Photosynthetic characteristics of rice leaves grown under red light with or without supplemental blue light. *Plant Cell Physiol.* 45, 1870–1874. doi: 10.1093/pcp/pch203
- McCree, K. J. (1971). The action spectrum, absorptance and quantum yield of photosynthesis in crop plants. *Agricult. Meteorol.* 9, 191–216. doi: 10.1016/0002-1571(71)90022-7
- Muggeo, V. M. R. (2003). Estimating regression models with unknown break-points. *Statistic. Med.* 22, 3055–3071. doi: 10.1002/sim.1545
- Naznin, M. T., Lefsrud, M., Gravel, V., and Azad, M. O. K. (2019). Blue light added with red LEDs enhance growth characteristics, pigments content, and antioxidant capacity in lettuce, Spinach, Kale, Basil, and sweet pepper in a controlled environment. *Plants* 8:93. doi: 10.3390/plants8040093
- Nogués, S., Allen, D. J. D., Morison, J. J. I. L., Baker, N. N. R., Nogués, S., Allen, D. J. D., et al. (1998). Ultraviolet-B radiation effects on water relations, leaf development, and photosynthesis in droughted pea plants. *Plant Physiol.* 117, 173–181. doi: 10.1104/pp.117.1.173
- Ouzounis, T., Rosenqvist, E., and Ottosen, C. O. (2015). Spectral effects of artificial light on plant physiology and secondary metabolism: a review. *HortScience* 50, 1128–1135. doi: 10.21273/hortsci.50.8.1128
- Pattison, P. M., Tsao, J. Y., Brainard, G. C., and Bugbee, B. (2018). LEDs for photons, physiology and food. *Nature* 563, 493–500. doi: 10.1038/s41586-018-0706-x
- Pennisi, G., Blasioli, S., Cellini, A., Maia, L., Crepaldi, A., Braschi, I., et al. (2019). Unraveling the role of red: blue LED lights on resource use efficiency and nutritional properties of indoor grown sweet Basil. *Front. Plant Sci.* 10:305. doi: 10.3389/fpls.2019.00305
- Poorter, H., Niinemets, U., Ntagkas, N., Siebenkas, A., Maenpaa, M., Matsubara, S., et al. (2019). A meta-analysis of plant responses to light intensity for 70 traits ranging from molecules to whole plant performance. *New Phytol.* 223, 1073–1105. doi: 10.1111/nph.15754
- Rama Das, V. S. (2006). Photosynthesis. Regulation under varying light regimes. *Photosynthetica* 44:364. doi: 10.1007/s11099-006-0037-2
- Reynolds, S. G. (1970). Gravimetric meth of soil moisture determination. *J. Hydrol.* 11, 258–273.
- Robinson, S. A. (2001). Plant light stress. *Encycl. Life Sci.* 1, 1–5.
- Ruban, A. V. (2009). Plants in light. *Commun. Interact. Biol.* 889, 50–55. doi: 10.4161/cib.2.1.7504
- Schneider, C. A., Rasband, W. S., and Eliceiri, K. W. (2012). NIH Image to ImageJ: 25 years of image analysis. *Nat. Methods* 9, 671–675. doi: 10.1007/978-1-84882-087-6_9
- Schreiner, M., Mewis, I., Huyskens-Keil, S., Jansen, M. A. K., Zrenner, R., Winkler, J. B., et al. (2012). UV-B-induced secondary plant metabolites – potential benefits for plant and human health. *Crit. Rev. Plant Sci.* 31, 229–240. doi: 10.1080/07352689.2012.664979
- Son, K. H., Jeon, Y. M., and Oh, M. M. (2016). Application of supplementary white and pulsed light-emitting diodes to lettuce grown in a plant factory with artificial lighting. *Horticult. Environ. Biotechnol.* 57, 560–572. doi: 10.1007/s13580-016-0068-y
- Son, K. H., and Oh, M. M. (2013). Leaf shape, growth, and antioxidant phenolic compounds of two lettuce cultivars grown under various combinations of blue and red light-emitting diodes. *HortScience* 48, 988–995. doi: 10.21273/hortsci.48.8.988
- Steyn, W. J., Wand, S. J. E., Holcroft, D. M., and Jacobs, G. (2002). Anthocyanins in vegetative tissues: a proposed unified function in photoprotection. *New Phytol.* 155, 349–361. doi: 10.1046/j.1469-8137.2002.00482.x
- Stutte, G. W., and Edney, S. (2009). Photoregulation of bioprotectant content of red leaf lettuce with light-emitting diodes. *HortScience* 9, 79–82.
- Takahashi, S., and Badger, M. R. (2011). Photoprotection in plants: a new light on photosystem II damage. *Trends Plant Sci.* 16, 53–60. doi: 10.1016/j.tplants.2010.10.001
- Taulavuoria, K., Hyökya, V., Oksanen, J., Taulavuoria, E., and Julkunen-Tiittob, R. (2016). Species-specific differences in synthesis of flavonoids and phenolic acids under increasing periods of enhanced blue light. *Environ. Exp. Bot.* 121, 145–150. doi: 10.1016/j.envexpbot.2015.04.002
- Terashima, I., Fujita, T., Inoue, T., Chow, W. S., and Oguchi, R. (2009). Green light drives leaf photosynthesis more efficiently than red light in strong white light: revisiting the enigmatic question of why leaves are green. *Plant Cell Physiol.* 50, 684–697. doi: 10.1093/pcp/pcp034
- Touliatos, D., Dodd, I. C., and Mcainsh, M. (2016). Vertical farming increases lettuce yield per unit area compared to conventional horizontal hydroponics. *Food Energy Secur.* 5, 184–191. doi: 10.1002/fes3.83
- Tsormpatsidis, E., Henbest, R. G. C., Battey, N. H., and Hadley, P. (2010). The influence of ultraviolet radiation on growth, photosynthesis and phenolic levels of green and red lettuce: Potential for exploiting effects of ultraviolet radiation in a production system. *Ann. Appl. Biol.* 156, 357–366. doi: 10.1111/j.1744-7348.2010.00393.x
- Wang, L. S., and Stoner, G. D. (2008). Anthocyanins and their role in cancer prevention. *Cancer Lett.* 269, 281–290. doi: 10.1016/j.canlet.2008.05.020
- WHO-FAO (2004). Fruit and vegetable for health: report of a joint FAO/WHO workshop, 1-3 september 2004, Kobe, Japan. *Handbook Plant Food Phytochem.* 4, 105–137. doi: 10.1002/9781118464717.ch5
- Wickham, H., Chang, W., Hermy, L., Pedersen, T. L., Takahashi, K., Wilke, C., et al. (2020). *Package 'ggplot2'*. Available Online at: <https://CRAN.R-project.org/package=ggplot2> (accessed June 19, 2020).
- Wilcox, J. K., Catignani, G. L., and Lazarus, C. (2003). Tomatoes and cardiovascular health. *Food Sci.* 43, 1–18.
- Zhang, Y., Xu, S., Cheng, Y., Peng, Z., and Han, J. (2018). Transcriptome profiling of anthocyaninrelated genes reveals effects of light intensity on anthocyanin biosynthesis in red leaf lettuce. *PeerJ* 2018, 1–18. doi: 10.7717/peerj.4607
- Zhu, H., Zhang, T. J., Zheng, J., Huang, X. D., Yu, Z. C., and Peng, C. L. (2018). Anthocyanins function as a light attenuator to compensate for insufficient photoprotection mediated by nonphotochemical quenching in young leaves of *Acmena acuminatissima* in winter. *Photosynthetica* 56, 445–454. doi: 10.1007/s11099-017-0740-1

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2020 Cammarisano, Donnison and Robson. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.