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Inducible glutathione S-transferase (*IrGST1*) from the tick *Ixodes ricinus* is a haem-binding protein

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Abstract

Blood-feeding parasites are inadvertently exposed to high doses of potentially cytotoxic haem liberated upon host blood digestion. Detoxification of free haem is a special challenge for ticks, which digest haemoglobin intracellularly. Ticks lack a haem catabolic mechanism, mediated by haem oxygenase, and need to dispose of vast majority of acquired haem *via* its accumulation in haemosomes. The knowledge of individual molecules involved in the maintenance of haem homeostasis in ticks is still rather limited. RNA-seq analyses of the *Ixodes ricinus* midguts from blood- and serum-fed females identified an abundant transcript of *glutathione S-transferase (gst)* to be substantially up-regulated in the presence of red blood cells in the diet. Here, we have determined the full sequence of this encoding gene, *ir-gst1*, and found that it is homologous to the delta-/epsilon-class of GSTs. Phylogenetic analyses across related chelicerates revealed that only one clear *IrGST1* orthologue could be found in each available transcriptome from hard and soft ticks. These orthologues create a well-supported clade clearly separated from other ticks' or mites' delta-/epsilon-class GSTs and most likely evolved as an adaptation to tick blood-feeding life style. We have confirmed that *IrGST1* expression is induced by dietary haem(oglobin), and not by iron or other components of host blood. Kinetic properties of recombinant *IrGST1* were evaluated by model and natural GST substrates. The enzyme was also shown to bind haemin *in vitro* as evidenced by inhibition assay, VIS spectrophotometry, gel filtration, and affinity chromatography. In the native state, *IrGST1* forms a dimer which further polymerises upon binding of excessive amount of haemin molecules. Due to susceptibility of ticks to haem as a signalling molecule, we speculate that the expression of *IrGST1* in tick midgut functions as intracellular buffer of labile haem pool to ameliorate its cytotoxic effects upon haemoglobin intracellular hydrolysis.

1. Introduction

Ticks are blood-feeding ectoparasites notorious for transmitting a wide variety of infection diseases of humans as well as farm and companion animals (de la Fuente et al., 2008). Hard ticks (Ixodidae) undergo a life cycle of three parasitic stages - larvae, nymphs, and adults, each of which requires one blood meal as the only source of nutrients for their further development and reproduction. Adult hard tick females imbibe large quantities of host blood exceeding up to hundred times their unfed weight. The blood meal is ultimately processed into the huge clutch of eggs before the female dies (Sonenshine and Roe, 2014). The proteinaceous components of the blood meal are internalised by tick digest cells lining up the midgut epithelium. The host proteins are then hydrolysed intracellularly, in the endo-lysosomal system consisting of a network of acidic cysteine and aspartic peptidases (Sojka et al., 2013). Haemoglobin degradation is inevitably concomitant with the intracellular release of haem, a pro-oxidative molecule, which is potentially cytotoxic when in excess (Graca-Souza et al., 2006).

We have recently demonstrated that ticks lost genes encoding enzymes involved in both haem biosynthesis and haem degradation (Perner et al., 2016a). Instead, ticks acquire haem exogenously, from the host haemoglobin (Perner et al., 2016a). A small portion of acquired haem is further dispatched for systemic inter-tissue distribution to allow assembly of endogenous haemoproteins, while most of the haem has to be disposed by effective means of detoxification. In contrast to haemozoin formation, a well-described mechanism of haem disposal in malaric plasmodium, schistosomes, or rhodnius vectors, ticks accumulate excessive haem as non-crystalline aggregates, in a specialised organelles generally referred to as haemosomes (Lara et al., 2005; Lara et al., 2003). Haem intracellular transport from digestive vesicles to cytosol was reported to be mediated by an ATP-binding cassette (ABCB10) (Lara et al., 2015). However, the next fate of cytosolic haem is still poorly understood.

In order to contribute to the knowledge of haem metabolism in ticks, we have tested, by RNA-seq analyses, which transcripts change their levels in response to the presence of red blood cells (RBCs) in the tick diet. For this purpose, we compared the transcriptomes of midguts from *I. ricinus* females membrane-fed either bovine blood or bovine serum. Among the surprisingly low number of transcripts with decreased or elevated levels in response to RBCs presence, we identified a gut-specific transcript Ir-114935 encoding a delta-/epsilon-class glutathione S-transferase (Perner et al., 2016b).

Members of the glutathione S-transferases (GSTs) family are ubiquitously present in eukaryotic organisms where they serve mainly in cellular detoxification of endogenous or xenobiotic compounds *via* their conjugation with the reduced glutathione (GSH), which results in their increased water solubility and excretion (Townsend and Tew, 2003; Wilce and Parker, 1994). Based on their organismal origin, primary sequence, substrate specificity, immunological, or chromosomal localisations, the GSTs can be grouped into more than a dozen classes historically tagged by Greek letters (Mashiyama et al., 2014). The

availability of the tick *Ixodes scapularis* genome (Gulia-Nuss et al., 2016) made it possible to enumerate and classify GSTs encoded in this species (Reddy et al., 2011). Out of 35 identified *IsGST* genes, 32 encoded cytosolic GSTs comprising 14 genes of vertebrate/mammalian Mu-class, 7 genes of Delta- and 5 genes of Epsilon- classes specific for insects, and 3 genes each were of common Omega- and Zeta- classes (Reddy et al., 2011). Given their capability to detoxify xenobiotics, GSTs have a well-established role in development of insecticide resistance in insect pests (for review see e.g. (Enayati et al., 2005) or acaricide resistance in ticks (Dreher-Lesnick et al., 2006; Duscher et al., 2014; He et al., 1999; Vaz et al., 2004a)). However, much less is known about the house-keeping physiological function of GSTs in the management of potentially toxic endogenous haem originating from the blood meal diet of the haematophagous parasites.

In this work, we provide a biochemical and functional characterisation of the haem(oglobin)-inducible GST from *I. ricinus* (further referred to as *IrGST1*) and demonstrate that this enzyme is capable to efficiently bind haemin *in vitro*. Clear orthologues of *IrGST1* could be found only in other tick species, but not in other organisms, suggesting that *IrGST1* represents a novel class of tick-specific GSTs that evolved during the adaptation of ticks to their blood-feeding life style.

2. Materials and methods

2.1. Tick maintenance and natural feeding

Adult *I. ricinus* ticks were collected in the forest near České Budějovice. Ticks were kept at 24 °C and 95% humidity under a 15:9-hour day/night regime. All laboratory animals were treated in accordance with the Animal Protection Law of the Czech Republic No. 246/1992 Sb., ethics approval No. 357 095/2012. The study was approved by the Institute of Parasitology, Biology Centre of the Czech Academy of Sciences (CAS) and Central Committee for Animal Welfare, Czech Republic (protocol no. 1/2015).

2.2. Tick membrane feeding

Membrane feeding of ticks was performed using a stationary six-well plate format according to Thomas Kröber and Patrick Guerin (Kröber and Guerin, 2007). Whole bovine blood was collected in a local slaughter house and manually defibrinated. To obtain serum, whole blood samples were centrifuged at $2500 \times g$, 10 min, 4°C and the resulting supernatant was collected and centrifuged again at $10\,000 \times g$, 10 min, 4°C. Fifteen females were placed in a feeding unit lined with a thin (80–120 µm) silicone membrane, previously pre-treated with a bovine hair extract in dichloromethane (0.5 mg of low volatile lipids). After 24 hr, unattached or dead females were removed and an equal number of males were added to the attached females into the feeding unit. Diets were exchanged in a 12h regime, with concomitant

addition of 1 mM adenosine triphosphate (ATP) and gentamicin (5 µg/ml). For diet supplementation, pure bovine haemoglobin (Sigma - H2500), bovine holo-Transferrin (Rocky Mountain Biologicals), and haemin (Sigma - H9039) was used. For membrane feeding experiments, haemin stock solution (62.5 mM haemin dissolved in 100 mM NaOH), was diluted 100× to reach 625 µM (final concentration) equalling a haem equimolarity with 1% haemoglobin (w/v).

2.3. Tissue dissection, RNA extraction, cDNA synthesis, and RT-qPCR

Membrane-fed *I. ricinus* females were forcibly removed from the membrane on day 6 of feeding. Tick midguts were dissected on a paraplast-filled Petri dish under a drop of ice-cold DEPC-treated PBS. Total RNA was isolated from dissected tissues using a NucleoSpinRNA II kit (Macherey-Nagel, Germany), quality checked by gel electrophoresis on agarose gel, and stored at -80°C prior to cDNA synthesis. cDNA preparations were made from 0.5 µg of total RNA in independent triplicates using the Transcriptor High-Fidelity cDNA Synthesis Kit (Roche Diagnostics, Germany). The cDNA served as templates for subsequent quantitative expression analyses by RT-qPCR. Samples were analysed by a LightCycler 480 (Roche) using Fast Start Universal SYBR Green Master Kit (Roche). Relative expressions were calculated using the $\Delta\Delta C_t$ method (Pfaffl, 2001). The expression profiles were normalised to *I. ricinus* elongation factor 1 α (*ef-1 α*). List of primers is available as Supplementary Table S1.

2.4. Sequencing, cloning, and phylogenetic analysis of *IrGST1*

Full cDNA sequence of gene encoding *IrGST1* was amplified using primers derived from 5'- and 3'- untranslated regions of the orthologous *I. scapularis* gene (ISCW005803) (Supplementary Table S1). *I. ricinus* midgut-specific cDNA prepared from midguts of females fed for 3 days served as template. The amplified 786 bp long PCR product was cloned into a pCR4-TOPO TA vector (Invitrogen) and sequenced. Amino acid sequences of *IrGST1* and other selected delta-/epsilon-class GSTs were aligned using the E-INS-i algorithm in MAFFT v7.017 (Katoh et al., 2002) and manually trimmed in Geneious v8.1.3. (Kearse et al., 2012) to the final length of 221 amino acids. Maximum parsimony analysis was performed in PAUP* v4.b10 (Swofford, 2003) using a heuristic search with random taxa addition, the ACCTRAN option, TBR swapping algorithm, all characters treated as unordered and gaps treated as missing data. Maximum-likelihood analysis was performed in RAxMLv7.2.8 (Stamatakis, 2006) under the PROTGAMMABLOSUM62 + Γ model. Mosquitoes *Anopheles gambiae* and *Aedes aegypti* were used as outgroups. Bootstraps were based on 1000 replicates for both analyses. Bayesian inference analysis was performed in MrBayes v3.0 (Ronquist and Huelsenbeck, 2003) using the WAG + Γ model of evolution. Analyses were initiated with random starting trees, four simultaneous MCMC chains sampled at intervals of 200 trees and posterior probabilities estimated from 1 million generations (burn-in 100 000).

2.5. Expression and purification of recombinant polyhistidine-tagged and untagged *IrGST1*

Poly-histidine tagged recombinant *IrGST1* was prepared by amplification of a coding sequence using *ir-gst1*-specific primers (Supplementary Table S1) and the 694 bp product was cloned into a pet100 vector (Invitrogen) following manufacturers' protocol. Resulting plasmid was transformed into TOP10 *E. coli* cells and positive clones were selected on ampicillin LB plates, sequenced, and transformed into *E. coli* BL21 Star (DE3) chemically competent cells (Invitrogen). Cells were grown in ampicillin LB medium at 37°C and when reached OD = 1.6, the culture was induced with 0.1 mM isopropyl β-D-1-thiogalactopyranoside (IPTG). The cells were cultured for 6 h and the harvested cells were suspended in PBS and homogenised using sonication. The centrifuged homogenate was loaded on a Ni²⁺-IMAC agarose (GE Healthcare) and the resin was washed with 20 bed volumes of 20 mM phosphate buffer pH 6.0, 0.5 M NaCl, 20 mM imidazol, 10% glycerol and 0.5% Triton X-100 (v/v) to remove non-specifically bound proteins. The recombinant protein was then eluted from the resin by 100 mM imidazole in a washing buffer, concentrated and transferred to the PBS by ultrafiltration using 15 ml centrifuge filter units (cut-off 10 kDa, Merck Millipore). The monospecific polyclonal antibodies against *IrGST1* were raised in rabbits as previously described (Grunclova et al., 2006) and used for Western blotting. A PCR product encoding an untagged recombinant *IrGST1* was amplified using the same primer pair (with the reverse primer containing a stop codon), cloned into the pet101 vector (Invitrogen), and further expressed in *E. coli* as described above. The cell pellet was homogenised in a tenth of original culture volume in 20 mM Tris pH 8.5 and centrifuged at 10 000 × g, 20 min, 4°C. The supernatant was filtered through a 0.22 μm filter (Merck Millipore) and applied on MonoQ HR 5/5 column (GE Healthcare) using an AKTA pure chromatographic system (GE Healthcare). The sample was separated at 1 ml/min flowrate in 20 mM Tris pH 8.5 and eluted by a 0–500 mM NaCl gradient. The fractions with enriched GST activity (see below) were pooled, concentrated by ultrafiltration (10 kDa cut-off), applied onto Superdex 75 10/300 GL column (GE Healthcare) and further separated at 1 ml/min flowrate in 20 mM Tris pH 8.5, 150 mM NaCl. Gel filtration molecular standards (ferritin, aldolase, serum albumin, and chymotrypsinogen A) were used for molecular weight determination. Purified *IrGST1* was used for immunising mice and the mice immune sera were used for immunohistochemistry (see below).

2.6. Reducing SDS-PAGE and Western blot analysis

Tick midgut homogenates were prepared in 1% Triton X-100 in PBS supplemented with a Complete™ cocktail of protease inhibitors (Roche) using a 29G syringe needle, and subsequently subjected to three freeze/thaw cycles using liquid nitrogen. Proteins were then extracted for 1 hr at 4°C and 1 200 rpm using a Thermomixer comfort (Eppendorf, Germany). Samples were then centrifuged 15 000 × g, 10 min, 4°C and separated by reducing SDS-PAGE on 12.5 % polyacrylamide gels. Protein profiles were visualized

using TGX stain-free chemistry (BioRad) or by staining with Coomassie Brilliant Blue R-250 (CBB). Proteins were transferred onto Immobilon-P[®] LF PVDF membrane using a Trans-Blot Turbo system (BioRad). For Western blot analyses, membranes were blocked in 3% (w/v) non-fat skimmed milk in PBS with 0.05% Tween 20 (PBS-T), incubated in immune serum (α IrGST1) diluted in PBS-T (1:5000) or in immune serum against *I. ricinus* ferritin 1 (α IrFer1) diluted in PBS-T (1:50). The goat anti-rabbit IgG-peroxidase antibody (Sigma A9169) diluted in PBS-T (1:50000) was used as a secondary antibody. Signals were detected using ClarityWestern ECL substrate, visualized using a ChemiDoc MP imager, and analysed using Image Lab Software (BioRad).

2.7. Substrate specificities of IrGST1 and isoelectric focusing

The substrate specificities of IrGST1 were tested with known model and natural substrates of GSTs (Morphew et al., 2012). In brief, enzyme assays were measured using a UV/VIS Gilford Response spectrophotometer over 3 min at 25°C. Glutathione S-transferase (GST) activity was determined using the model substrate 1-chloro-2,4-dinitrobenzene (CDNB) according to the method described by Habig et al. (Habig et al., 1974). The enzyme assay was performed in 100 mM potassium phosphate buffer pH 6.5, containing 1mM reduced glutathione (GSH) and 1mM CDNB at 340 nm ($\epsilon = 9.6 \times 10^6 \text{ cm}^2 \text{ mol}^{-1}$). GSH-dependent peroxidase activity of IrGST1 was determined using cumene hydroperoxide (the model lipid hydroperoxide substrate)(Jaffe and Lambert, 1986). The assay was carried out in 50 mM phosphate buffer pH 7.0 containing 1 mM GSH, 0.2 mM NADPH, 0.5 U glutathione reductase (Sigma, G3664), and 1.2 mM cumene hydroperoxide. The reaction was measured at 340 nm ($\epsilon = 6.22 \times 10^6 \text{ cm}^2 \text{ mol}^{-1}$). GST activity with trans-2-nonenal was determined as previously described (Brophy et al., 1989) The reaction mix was composed of 100 mM potassium phosphate buffer pH 6.5, 1 mM GSH, and 0.23 mM trans-2-nonenal and the enzymatic activity was measured at 225 nm ($\epsilon = -19.2 \times 10^6 \text{ cm}^2 \text{ mol}^{-1}$). Isoelectric focusing (IEF) was performed on IEF precast gels (BioRad) and separated by IEF cathode buffer 2mM lysine (free base), 2 mM arginine (free base) and IEF anode buffer 0.7 mM phosphoric acid at increased voltage modes: 100 V 60 min, 250 V 60 min, 500 V 30 min. IEF markers of pI range 3–10 were used (SERVA 39212.01).

2.8. Determination of kinetic parameters and inhibition studies

The determination of IrGST1 apparent Michaelis constants to CDNB and GSH as substrates were performed in triplicates, with varying concentrations of CDNB and constant GSH (1 mM), or constant CDNB (1 mM) and varying concentrations of GSH, respectively. Kinetic constants were calculated by non-linear regression analysis of the experimentally measured activities. Data were fitted to the Michaelis-Menten equation using GraphPad Prism 6.0 software. The inhibition of IrGST1 by haem-related

compounds was investigated by CDNB activity assay with haemin (haem-chloride), haematin (haem-hydroxide, Sigma H3281), protoporphyrin IX (PPIX) (Sigma P8293), and myoglobin (Sigma M0630). Haemin and haematin were dissolved in 100 mM NaOH, PPIX in DMSO, and myoglobin in H₂O.

2.9. Examination of haemin-*IrGST1* binding by spectrophotometry and haemin-affinity chromatography

The absorption properties of haemin in the presence of *IrGST1* were measured by recording the absorption spectra in the range from 300 nm to 450 nm using a UV-1800 spectrophotometer (Shimadzu). Constant concentration of haemin (10 µM) in 20 mM sodium phosphate buffer (pH 8.0), 50 mM NaCl was incubated for 15 min with different concentrations of *IrGST1* corresponding to the molar ratios of *IrGST1*: haemin from 0 to 2.

Binding of *IrGST1* to the commercial haemin-agarose (Sigma H6390) was examined using a pull down experiment. *E. coli* expressing *IrGST1* (see above) were homogenised in 1.5 ml of 50 mM Tris-HCl pH 8.0, 0.5 M NaCl (Tris-NaCl buffer), centrifuged and the supernatant was incubated with 50 µl of haemin-agarose for 1 h with slow rotation. Agarose beads were then allowed to settle, supernatant removed, and the agarose was then transferred to an empty column (BioRad) and extensively washed with Tris-NaCl buffer. Specifically bound proteins were then eluted with 1 M urea in Tris-NaCl buffer and separated using SDS-PAGE. Coomassie-stained protein band was excised and prepared for mass fingerprint analysis. Briefly, the excised gel was incubated with 50 mM ammonium bicarbonate: 100% acetonitrile (1:1) solution for 15 minutes at 37°C to destain the gel. The gel was dehydrated in 100% acetonitrile for 30 minutes at 37°C and subsequently rehydrated in trypsin solution (100 ng/µl) in ammonium bicarbonate, left for 45 minutes at 8°C, and incubated at 37°C over-night. Supernatant was removed and the gel was washed several times alternately with acetonitrile and 50 mM ammonium bicarbonate. Samples were then vacuum-dried, resuspended 1% formic acid (w/w) and analysed by LC MS/MS on an Agilent 6550 iFunnel Q-TOF mass spectrometer with a Dual AJS ESI source coupled to a 1290 series HPLC system (Agilent, Cheshire, UK) according to Morphew et al. 2014 (Morphew et al., 2014).

2.10. Haemin binding size exclusion chromatography

Haemin binding size exclusion chromatography was performed with *IrGST1*, diluted to 35 µM (0.9 mg/ml) in 20 mM Tris pH 8.5, 150 mM NaCl. Different molar ratios of haemin (stock solution 35 mM haemin in 100 mM NaOH) were added to 1 ml of *IrGST1* solution and incubated for 30 minutes, 250 rpm, 22°C in ThermoMixer® (Eppendorf). Then 250 µl of the membrane filtered (0.22 µm) incubation reaction was separated using Superdex 75 10/300 GL column (GE Healthcare) at 1 ml/min flow rate in 20 mM Tris pH 8.5, 150 mM NaCl and the absorbance was monitored at the dual wavelengths of 280 and 400 nm.

2.11. RNA interference and immunohistochemistry

dsRNA of *ir-gstI* or *gfp* (green fluorescent protein) used for control were synthesised using the MEGAscript T7 transcription kit (Ambion, Lithuania) according to the previously described protocol (Hajdušek et al., 2009). *I. ricinus* females were injected into the haemocoel through to the coxae with *ir-gstI*-specific dsRNA or control *gfp* dsRNA (0.5 µl; 3 µg/µl) using a microinjector (Narishige), allowed to rest for one day and then fed naturally for 5 days on guinea pigs. The efficiency of RNA-mediated silencing *ir-gstI* gene expression was verified at the protein level by Western blot analysis using $\alpha IrGST1$ antibodies. The visualisation of authentic *IrGST1* by indirect immune-fluorescent microscopy in the dissected *I. ricinus* guts was performed as described earlier (Franta et al., 2010) with some modification. Briefly, the semi-thin sections were cut, transferred onto glass slides, and blocked with 1% BSA and 10% goat serum PBS-T (0.3% Triton X-100) for 1 h. Incubation with the primary $\alpha IrGST1$ antibody (1:100) in PBS-T was performed in a humid chamber for 1.5 h at room temperature. For negative control experiments, the primary antibody incubation was omitted (not shown). Sections were washed with PBS-T (four times 5 min) and then incubated with Alexa Fluor® 488 secondary dye-conjugated goat anti-mouse (Invitrogen/Molecular Probes) diluted to 1:500 in PBS-T for 1 h at room temperature. After washing with PBS-T, the slides were counterstained with DAPI (4',6'-diamidino-2-phenylindole; 2.5 µg/ml; Sigma) for 5 min. Finally, sections were mounted in Fluoromount (Sigma-Aldrich) and examined using the fluorescence microscope BX 53. The semi-thin sections stained with toluidine blue were examined under the light mode of the microscope BX53. The same protocol was carried out for immunodetection of 4-hydroxynonenal (4-HNE) using the commercial rabbit $\alpha 4$ -HNE antibody (Abcam ab46545, 1:300) and the 1:500 diluted Alexa Fluor® 488-labeled secondary goat anti-rabbit antibody (Invitrogen/Molecular Probes).

3. Results

3.1. *IrGST1* sequence identification and phylogenetic analysis

The transcript Ir-114935, previously shown to be significantly up-regulated in the midguts of blood-fed compared to serum-fed ticks (Perner et al., 2016b), encodes a partial sequence of a putative delta-/epsilon-class glutathione S-transferase. The partial sequence was clearly orthologous to *I. scapularis* ISCW005803 gene encoding delta-class GST, *IsGSTD2* (Reddy et al., 2011), with 98% sequence identity at both amino-acid and nucleotide levels. The full coding sequence of *IrGST1* (deposited in the GenBank under Access. No. MF984398) was obtained by cloning and sequencing of a 786 bp long PCR product amplified using the primers derived from 5'- and 3'-UTR regions of *I. scapularis* *IsGSTD2*. All performed phylogenetic analyses have unambiguously revealed the *IrGST1* orthologues of delta-/epsilon-

class from other hard and soft tick species that create a well-supported clade (Fig. 1A). This specific clade is distinct from the clades of other delta-/epsilon- class GSTs from *I. scapularis*, horseshoe crab, and mites (Fig. 1A), which display much lower sequence identity to *IrGST1* (below 50%) (Fig. 1B and Supplementary Fig. S1). The typical SNAIL/TRAIL signature motif present in GSTs of various classes is conserved within the *IrGST1* orthologous group as SRAI(A/G). All ticks and mites delta-/epsilon-class GSTs possess a conserved tyrosine residue at the position 6 (Fig. 1B and Supplementary Fig. S1) classifying the enzymes into Y-type major subgroup of cytosolic GSTs (Atkinson and Babbitt, 2009).

3.2. Expression of *IrGST1* in the tick gut is inducible by haemin

Even though we have previously demonstrated by RNA-seq and qRT-PCR that expression of contig Ir-114935 encoding *IrGST1* is up-regulated in ticks fed on the whole blood (Perner et al., 2016b), a question remaining to be solved was which constituent of red blood cells is responsible for the up-regulation of the *ir-gst1* gene. In order to reveal that, we have conducted a membrane feeding experiment where the ticks were allowed to feed for 5 days haemoglobin-free serum, haemoglobin-free serum supplemented either with 1% w/v bovine haemoglobin, 625 μ M haemin (haemin solubilised in sodium hydroxide), mock (sodium hydroxide, 1 mM final concentration), whole blood reconstituted with red blood cells. RT-qPCR analysis revealed that *ir-gst1* is up-regulated by dietary haemin as well as by haemoglobin (Fig. 2A). The up-regulation was observed higher for dietary haemin (unbound or secondarily complexed with albumin) compared to dietary haemoglobin, when the concentration of haem were equimolar. The consistent result was obtained at the protein level, as evidenced by Western blot analysis (Fig. 2B). The levels of *IrGST1* in the midgut were dose-dependent on the amount of haemin added to the serum diet and gradually increased from micromolar dietary concentration to 625 μ M representing about 1/150 of the physiological concentration of haem present in the whole blood ($\sim$ 10 mM) (Fig. 2C). This result indicates that *IrGST1* inducibility by host haem may serve as a very sensitive sensor of the blood meal uptake. To confirm that *IrGST1* expression is induced exclusively by haem and not by iron, we performed a membrane feeding experiment where ticks were fed diets enriched with bovine holo-transferrin and bovine haemoglobin, known tick sources of iron and haem, respectively (Perner et al., 2016a). We confirmed that while ticks fed transferrin-enriched diet had elevated levels of *IrFer1*, *IrGST1* levels remained unaltered. Conversely, ticks fed haemoglobin enriched diet had elevated levels of *IrGST1* and unaltered levels of *IrFer1* (Fig. 2D). These results underscore that haem and iron sensing in the tick midgut are independent processes.

3.3. RNAi effectively silences the expression of *IrGST1* throughout the tick feeding

To study a physiological role of *IrGST1* *in vivo*, a knock-down of this transcript was obtained by RNAi-mediated silencing. A confirmation of clear down-regulation of *IrGST1* in tick midgut throughout the

feeding was evidenced by Western Blotting (Fig. 3A). Indirect immune-fluorescence microscopy using α *IrGST1* antibody confirmed a substantial decrease of fluorescent signal in the matured midgut digest cells of *IrGST1*-KD females compared to the control ticks injected with *gfp* dsRNA (Fig. 3B). However, this reduction of *IrGST1* in tick midgut digest cells had no obvious impact on the tick feeding and fecundity. *IrGST1*-KD females could accomplish feeding, reached comparable engorged weights, laid egg clutches of comparable size and colour that gave rise to viable larvae similarly as the control *gfp* group (Fig. 3C, D). GSTs are also known to conjugate GSH to 4-hydroxynonenal (4-HNE), that is a toxic product of lipid peroxidation and a biomarker of oxidative stress (Awasthi et al., 2004; Cheng et al., 2001). The presence of 4-HNE inside the digest cells was detected by indirect immune-fluorescent microscopy. However, no difference of fluorescent intensity was observed in guts from *IrGST1*-KD and control females (Supplementary Fig. S2), suggesting that *IrGST1* is not involved in 4-HNE detoxification. Given the absence of any *IrGST1*-KD phenotype, we further focused on *in vitro* characterisation of recombinant *IrGST1*.

3.4. Preparation and enzymatic characterisation of recombinant *IrGST1*

Recombinant *IrGST1* was first expressed in *E. coli* expression system as a His-tagged protein with a theoretical mass 29721 Da and purified in a soluble form from the bacterial lysate using Ni^{2+} -IMAC chromatography under native conditions (Fig. 4A). *E. coli* expressed *IrGST1* was active in GSH transferase activity assay using a model substrate 1-chloro-2,4-dinitrobenzene (CDNB) (Fig. 4B). The crude extract of *E. coli* cells had a specific enzymatic activity of 201 ± 13 nmol CDNB/min/mg protein that following elution and dialysis increased to 1820 ± 77 nmol CDNB/min/mg protein in the purified fraction with K_m for GSH to be 0.87 ± 0.13 mmol and K_m for CDNB to be 2.9 ± 0.64 mmol (Supplementary Fig S3). Despite the capacity to utilise GSH in a CDNB activity assay (K_m values for GSH and CDNB were comparable to other reported GSTs (Al-Qattan et al., 2016)), the effort to purify *IrGST1* from *E. coli* crude extract or further purify the *IrGST1* (IMAC purified) using GSH- or S-hexyl GSH-Sepharose failed given the low binding affinity to these sorbents (Supplementary Fig. S3). GSTs display a wide range of enzymatic activities with model substrates (Brophy et al., 1990). To reveal the kinetic parameters of *IrGST1*, assays with typical GST substrates were also carried out using another invertebrate recombinant GST (*Fasciola gigantica* sigma GST) as a control (Morphew et al., 2012). Beside the above mentioned GSH-conjugating activity using CDNB as a model substrate, the *IrGST1* exerted also peroxidase activity with the model lipid hydroperoxide substrate - cumene hydroperoxide (Fig. 4B). *IrGST1* had no activity towards reactive carbonyls, in contrast to sigma-class *F. gigantica*, GST as assayed using trans-2-nonenal as a natural substrate derived from lipid peroxidation (Brophy et al., 1989) (Fig. 4B).

As several GSTs from different blood feeding parasites have been reported to bind haemin (see below), we have performed a haemin interaction/inhibitory assay of *IrGST1* GSH-conjugating activity and compared it to the *FgGST* over a range of CDNB substrate concentration (Fig. 4C). This experiment revealed that *IrGST1* activity is much more sensitive to haemin than *FgGST*. In order to determine more precisely the inhibition constant of haemin (K_i) for *IrGST1*, a Dixon plot of inhibitory activities was used and K_i of haemin was determined to be 42 ± 15 nM, indicating a strong binding affinity for *IrGST1* (Fig. 4D). Using the CDNB activity assay, we further examined whether free haem, protein-bound haem, or iron-free protoporphyrin IX (PPIX) is responsible for the inhibition of *IrGST1*. Both free haemin (haem-chloride) and haematin (haem-hydroxide) inhibited the *IrGST1* activity with apparent IC_{50} around 200 nM, whereas myoglobin-bound haem or iron-free protoporphyrin IX (PPIX) did not induce any inhibition at the concentration range up to 1 μ M (Fig. 4E).

3.5. Recombinant *IrGST1* binds haemin in vitro

To further support evidence of haemin binding to the *IrGST1*, crude extract from *E. coli* expressing *IrGST1* was incubated with haemin-agarose in a pull down assay. Roughly 75 % of *E. coli* extract proteins remained unbound in the supernatant, whereas the CDNB-specific activity was virtually lost (Fig. 5A), likely confined to the haemin-agarose pellet. After extensive washing with 20 mM phosphate buffer pH 7.4, 0.5 M NaCl, haemin-bound proteins were eluted with washing buffer supplemented with 1 M urea. SDS-PAGE analysis showed a major protein band of about 29 kDa (Fig. 5A), which was submitted to the peptide mass-fingerprint analysis that confirmed its identity as *IrGST1* with about 19% sequence coverage. To further elucidate the binding characteristics of haemin to *IrGST1*, we evaluated the binding properties by spectrophotometry at visible wavelength range. Unbound haemin displays an absorption maximum at $\lambda = 385$ nm, but its absorption maximum undergoes intensifying red-shifts upon binding to *IrGST1*, with isobestic point at $\lambda = 402$ nm, in the titration experiment where the *IrGST1* was treated as a ligand and haemin concentration was kept constant at 10 μ M (Fig. 5B). The point of saturation on the monitoring wavelength ($\lambda = 421$ nm) was reached at 1:1 molar ratio (Fig. 5C), indicating that one *IrGST1* molecule binds one molecule of haemin.

3.6. Native *IrGST1* is a haemin-binding dimer

In order to rule out a possible contribution of protein poly-histidine tag to haemin binding, untagged recombinant *IrGST1* was expressed in *E. coli* system and purified using two-step liquid chromatography by anion exchange chromatography (IEX) on the MonoQ column followed by size exclusion chromatography (SEC) on a Superdex 75 column (Fig. 6A). Monitoring absorbance at both $\lambda = 280$ nm and $\lambda = 400$ nm, revealed that the purified *IrGST1* already displayed some basal absorbance at 400 nm,

indicating that *IrGST1* strips endogenous haem from expressing *E. coli* cells (Fig. 6B). The molecular weight of the *IrGST1* was determined by SEC using calibration standards and calculated to be of 56.7 kDa implying a dimeric form (Fig. 6B). Isoelectric point of the native *IrGST1*, determined experimentally by isoelectric focusing, was pI 5.5 (Supplemental Fig. S4). To assess the molecular arrangement of the *IrGST1* - haemin complex, the *IrGST1* was titrated with different molar concentrations of haemin and the products were analysed by SEC. As shown in the Fig. 6C, *IrGST1* has the capacity to bind haemin as a dimer. When haemin is in excess, up to eight to one molar ratios to *IrGST1*, a fraction of the protein shifts towards higher molecular weights suggesting further polymerisation or aggregation of the *IrGST1*-haemin complex (Supplementary Fig. S5).

4. Discussion

The glutathione S-transferases (GSTs) from pathogens that carry out disposal of toxic endogenous and exogenous compounds have been investigated as potential targets for development of efficient anti-parasitic drugs and vaccines for three decades (Brophy et al., 1990; Harwaldt et al., 2002; Nare et al., 1992; Ricciardi and Ndao, 2015; Wei et al., 2016; Zhan et al., 2005; Zhan et al., 2010).

In this work, we have characterised a novel haem-inducible GST from the hard tick *I. ricinus* (*IrGST1*), which transcript was markedly up-regulated in blood-fed ticks compared to ticks fed with serum (Perner et al., 2016b). *IrGST1* belongs to the delta-/epsilon-class (insect type) GSTs and only one clear orthologue could be found among other 32 cytosolic GSTs identified in the genome of a closely relative species *I. scapularis* (Reddy et al., 2011). Accordingly, only one transcript orthologous to *IrGST1* could be found in transcriptomes available for other tick species. One haem-responsive GST, namely GST-19 (CE09995) has been identified using a proteomic approach among 36 nu-class members annotated in the genome of the model nematode *Caenorhabditis elegans* (Perally et al., 2008). Twenty six genes encoding GSTs were annotated in the mosquito *Aedes aegypti*, out of which the gene *gstx2* (new, unclassified class) showed an affinity for haematin (Lumjuan et al., 2007). Lately, a substantial up-regulation of this gene transcript was demonstrated using a transcriptome-wide analysis of haem influence on *A. aegypti* cell line (Bottino-Rojas et al., 2015).

Our phylogenetic analyses grouping haem-binding *IrGST1* with just one orthologue in each tick species (Fig. 1A, B) suggests that this tick-specific group likely exapted from an ancestral catalytic GST to form haem-binding GSTs under the evolution pressure exerted by their blood-feeding life style. This is supported by a distant phylogenetic positioning of tick putative haem-binding GSTs from other delta-/epsilon-class GSTs present in related non-haematophagous chelicerates (horseshoe crab and mites).

Several GSTs that have been at least partially characterised in ticks so far belong mostly to the mu-class (see (Shahein et al., 2013) for review). The first tick GST was purified from the larval stage of the cattle

tick *Rhipicephalus (Boophilus) microplus* using glutathione affinity chromatography (He et al., 1999). A gene encoding another mu-class GST from *R. microplus* (referred to as *BmGST*) was isolated from salivary gland cDNA library, and its tissue and developmental stage profiling revealed that *bm-gst* gene is transcribed in salivary glands and midguts of the adult females but not in the larval stage (Rosa de Lima et al., 2002). The enzyme kinetic parameters of the recombinant *BmGST* using the CDNB assay were determined together with the inhibitory potential for a number of compounds present in commercial acaricides (Vaz et al., 2004a). Two mu-class GSTs were cloned from the tick *Haemaphysalis longicornis* (*HlGST*) and *Rhipicephalus appendiculatus* (*RaGST*). The recombinant enzymes were reported to be effectively inhibited by acaricides, especially ethion and deltamethrin (Vaz et al., 2004b). Given the cross-reactivity of antibodies raised against recombinant *HlGST* from *H. longicornis* with *BmGST* from *R. microplus*, the cattle immunized with *HlGST* were partially protected against infestation by the cattle ticks (Parizi et al., 2011) that implies tick GSTs as a candidate antigen for anti-tick vaccine development (de la Fuente et al., 2016). Most recently, the vaccination potential of *HlGST* was tested against rabbit infestation with two closely related species *R. appendiculatus* and *R. sanguineus*. Despite the close similarity of GSTs from these tick species, the partial protection was obtained only against adult *R. appendiculatus* females (vaccine efficacy was reported to be of about 67%), but no protection was achieved against any stage of *R. sanguineus* (Sabadin et al., 2017). Another mu-class GST (tagged as *BaGST*) was cloned from the Egyptian cattle tick *R. (B.) annulatus* and the active recombinant *BaGST* was expressed and purified as fusion protein His-tagged protein (Shahein et al., 2008). This GST was later shown to be effectively inhibited by phenolic compounds and flavonoids from plant extracts but also by a haematin (Guneidy et al., 2014). RNAi-mediated silencing of the GST from *R. sanguineus* revealed the role of this enzyme in permethrin detoxification as the GST-KD ticks were more susceptible to the acaricide exposure (Duscher et al., 2014). The only partially characterised tick delta-/epsilon-class GST member was the *DvGST1* from *Dermacentor variabilis* (Fig. 1). The gene encoding *DvGST1* was reported to be specifically expressed in the tick gut and up-regulated by blood feeding (Dreher-Lesnick et al., 2006), which is suggestive to have a similar function as *IrGST1*. *Ir-gst1* expression is tissue-specific for *I. ricinus* midgut (Perner et al., 2016b), where it is localised to the cytosol of the digest cell (Fig. 3B). Its mRNA and protein levels are markedly up-regulated by addition of a soluble haemin into the diet. The inducibility is more sensitive for soluble haemin rather than for haem bound as a prosthetic group of haemoglobin (Fig. 2). This might indicate that free haemin is taken up by the digest cells by an alternative route independently of the proposed haemoglobin-specific receptor mediated uptake, potentially disbalancing intracellular haem homeostasis (Lara et al., 2005; Sojka et al., 2013). Importantly, haem sensing in the tick midgut is independent of iron sensing in the tissue, as *IrGST1* expression is induced only by dietary haem, and not by dietary iron (Fig. 2D). Unlike the iron

homeostasis, which is controlled at the translational level by proteosynthesis of intracellular iron storage protein - ferritin 1 (Hajdušek et al., 2009; Kopacek et al., 2003; Perner et al., 2016a), haem-inducible expression of *IrGST1* is apparently controlled at the level of gene transcription. In mammals, inducible GSTs are known to be regulated through antioxidant response element by the Keap1-Nrf2 pathway (Kansanen et al., 2013). Although several Nrf2-related transcripts containing basic leucine zipper (bZIP) domain have been found to be present in the *I. ricinus* midgut transcriptome (Perner et al., 2016b), their possible role as haem-responsive transcription factor(s) remains to be investigated.

Beside the *ir-gst1* inducibility by a dietary haem, we found that the GSH-conjugation activity of recombinant *IrGST1* (determined by CDNB assay) is noncompetitively inhibited by soluble haemin but not by iron-free porphyrin ring (protoporphyrinogen IX) or protein-bound haem (myoglobin) (Fig. 4). We further showed that, in the native state, *IrGST1* forms a dimer and binds haem in an equimolar ratio. The K_i of haemin to *IrGST1* was in the mid nanomolar range, comparable to the inhibition constants reported earlier for GSTs from other hematophagous parasites (Torres-Rivera and Landa, 2008). Haem-binding properties were investigated for several nematode-specific nu-class GSTs from different hookworm species such as *Haemonchus contortus*, *Ancylostoma caninum*, *Necator americanus*, or *Ancylostoma ceylanicum* (van Rossum et al., 2004; Wei et al., 2016; Zhan et al., 2005; Zhan et al., 2010). Recombinant *Na-GST1* from *N. americanus* elicited a significant reduction of hookworm burdens in vaccinated hamsters and hereby it became a leading vaccine candidate to prevent human hookworm infections (Zhan et al., 2010). Several haem-binding GSTs were also characterised in flatworms such as the sigma-class GSTs from flukes *Fasciola hepatica* (Brophy et al., 1990) or *FgGST-S1* from *F. gigantica* (Morphew et al., 2012). Much attention has been paid also to the haem-binding GSTs from different *Schistosoma* sp. (Walker et al., 1993) especially for their potential as promising vaccine candidates against human or bovine schistosomiasis (Capron et al., 2001; Capron et al., 1995; Ricciardi and Ndao, 2015). The most thoroughly investigated haem-binding GST has been *PfGST*, the only cytosolic GST encoded in the genome of the malaria parasite *Plasmodium falciparum* (Harwaldt et al., 2002; Liebau et al., 2002). The quite abundant *PfGST*, constituting about 3% of the total extractable proteins, forms in its native state a homodimer (Harwaldt et al., 2002) and was shown to capture haem that failed to be detoxified via polymerization in haemozoin (Liebau et al., 2002; Liebau et al., 2005). The 3D molecular structure of *PfGST* was resolved by X-ray diffraction (Fritz-Wolf et al., 2003) that further allowed to perform a molecular docking for a variety of *PfGST* ligands including haemin (Al-Qattan et al., 2016). We have demonstrated, using size exclusion chromatography (Fig. 6 C), that, in its native state, *IrGST1* binds haemin as a dimer at 1:1 molar ratio (Fig. 6 C). Moreover, the *in vitro* experiment showed that *IrGST1*-haemin binding occurs in the absence of reduced glutathione, a feature reported also for the *PfGST* (Liebau et al., 2009). In this work, the authors also described a transition of active *PfGST* dimer to an

inactive tetramer. We have observed a similar polymerisation of *IrGST1* in the presence of molar excess of haemin (Supplementary Fig. S5), suggesting that inhibition of *IrGST1* enzyme activity by haemin resulted also in formation of inactive polymer.

As mentioned above, several GSTs became a potential target for development of anti-parasitic vaccines (Capron et al., 2001; Capron et al., 1995; Parizi et al., 2011; Ricciardi and Ndao, 2015; Zhan et al., 2010). Although *IrGST1* seems to be the only haem-binding GST expressed in the tick gut, its RNAi-mediated silencing did not result in any clear phenotype impairing tick feeding and further development. Also our pilot vaccination experiments with recombinant *IrGST1* did not elicit any protection of immunised rabbits against tick infestation (data not shown). These findings rather limit the potential of *IrGST1* as a target for anti-tick intervention.

Under normal situation, ticks seem to be capable to efficiently detoxify excessive haem *via* its aggregation in haemosomes (Lara et al., 2003). Based on the functional analogy with the *P. falciparum* *PfGST* (Harwaldt et al., 2002; Liebau et al., 2009; Muller, 2015), our biochemical data indicates that *IrGST1* serves as a “back-up” guard molecule, i.e. by mopping up excess haemin and/or neutralising *via* lipid peroxidase activity the downstream consequence of haemin assault on membranes. Thus, *IrGST1* acts as ligandin, when high haemin levels override haemosome capacity and haemin is free in the cytosol and thus harmful to the tick.

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Legends to the Figures

Fig 1. Phylogeny and multiple alignment of selected chelicerate delta-/epsilon-class GST homologues. (A) Maximum likelihood phylogenetic tree of delta-/epsilon-class GST homologues in chelicerates with *IrGST1* and its tick orthologues grouping in one well-supported clade. Nodal supports are shown for maximum likelihood and maximum parsimony bootstraps and Bayesian inference posterior probability. Sequences used for multiple amino-acid alignment in Fig. 1B are shown in bold (B) Multiple amino-acid alignment of *IrGST1* sequence with putative orthologues from other ticks and selected delta-/epsilon-class GSTs from other chelicerates. **Hard ticks:** *I. ricinus* - *IrGST1* (this work, GenBank MF984398); *I. scapularis* - *IsGSTD2* (GenBank XM_002436245); *I. persulcatus* (GenBank GBXQ01020781); *R. appendiculatus* (GenBank GEDV01003209); *R. microplus* (GenBank GEFA01011362); *A. aureolatum* (GenBank GFAC01002707); *D. variabilis* *DvGST1* (GenBank AY241958). **Soft ticks:** *O. moubata* (GenBank GFJQ02000585); *C. mimon* (GenBank GEIB01001162). **Mites:** *D. gallinae* (GenBank KR337506); *G. occidentalis* (GenBank XM_003746739); *T. urticae* (GenBank XM_015936065); *S. scabiei* (GenBank AY649788); *V. destructor* (GenBank XP_022657236). **Horseshoe crab:** *L. polyphemus* (GenBank XM_013931267). The hash symbol points to the conserved tyrosine residue of Y-type GSTs. The asterisks and dashed red frame depict the conserved GSTs signature motif of *IrGST1* orthologues as SRAI(A/G). The column next to the sequences shows sequence identity percentage related to *IrGST1*.

Fig. 2. Analysis of *ir-gst1* expression by RT-qPCR and *IrGST1* levels by Western blotting. (A, B) Analysis of midguts of ticks fed serum for 5 days (S), serum supplemented with haemin solvent (S-mock), 625 μ M haemin (S+hm), 1% haemoglobin (equimolar to 625 μ M haemin) (S+Hb), or reconstituted blood

with 50% haematocrit (whole blood, WB). (A) RT-qPCR expression data are normalised against *elongation factor 1 α* (*ef1 α*). Shown data represent mean and SEM from three biological replicates. (B) Tick midgut homogenates were separated using reducing SDS-PAGE. Western Blot analysis was performed using specific rabbit anti-serum raised against *IrGST1* (α *IrGST1*) diluted 1:5000. (C) Midgut homogenates of ticks fed, for five days, serum supplemented with increasing haemin concentration were separated on reducing SDS-PAGE and Western Blot analysis was performed using α *IrGST1* (1:5000). (D) Midgut homogenates of seven days fed ticks, five days fed serum and for two consecutive days fed serum supplemented with 3 mg/ml bovine holo-Transferrin (Tf) or 10 mg/ml bovine Haemoglobin (Hb), were separated on reducing SDS-PAGE and Western Blot analysis was performed using α *IrGST1* (1:5000) and α *IrFer1* (1:50)

Fig. 3. RNAi verification and phenotypisation. Adult *I. ricinus* ticks were injected with *ir-gst1* or *gfp* (control) dsRNA and allowed to recover for 24 hours (Day 0). Ticks (n = 25) were then placed on a rabbit and allowed to feed naturally for indicated time-points (Day 3, Day 5, and Day 7), then forcibly removed, weighed out, and their midguts dissected (n $\geq$ 3). The remaining ticks were allowed to feed until repletion. (A) Tick midgut homogenates were separated by reducing SDS-PAGE and *IrGST1* protein levels were analysed by Western blotting using α *IrGST1* antibody (1:5000). (B) Sections were prepared from guts dissected from semi-engorged *I. ricinus* females (5 days of feeding). Section were labelled with α *IrGST1* serum (1:100) and Alexa488-conjugated anti-mouse antibody (left). DAPI was used to counterstain the nuclei. Sections were also stained with toluidine blue (right) - general structure of the tick gut showing the boundary between the gut epithelium and the gut lumen, containing large haemoglobin crystals (Hb) and developed digest gut cells (dGC); Nc - nuclei. (C) Column graph of tick weights before feeding and from individual time-points of feeding. Mean and SEM are shown, n $\geq$ 3. (D) Oviposition and larvae hatching from fully engorged females upon RNAi.

Fig. 4. Purification of recombinant *IrGST1*, substrate profiling, and haemin inhibition assay. (A) Soluble recombinant (His)₆-tagged *IrGST1* was purified from the lysate of expressing *E. coli* using Ni²⁺-IMAC under native conditions and fractions were analysed by reducing SDS-PAGE. Crude – *E. coli* lysate; Flow-through – unbound proteins; Elution – *IrGST1* eluted with 200 mM imidazole. (B) *IrGST1* from *I. ricinus* was tested for specific activities in spectrophotometric assays using GST model substrates; recombinant GST from *Fasciola gigantica* was used as a positive control (Morphew et al., 2012); 1-chloro-2,4-dinitrobenzene (CDNB) was used to test GSH-conjugation activity, trans 2-nonenal (T2N) was used to test GST-mediated reactions with reactive carbonyls, cumene hydroperoxide (C-HPx) was used to test peroxidase activity (C) Inhibition assays of haemin on CDNB activity of *IrGST1* and *F. gigantica*

over a range of different CDNB concentrations. **(D)** Dixon plot of inhibitory properties of haemin on *IrGST1* activity under different substrate (CDNB) concentrations. Calculated inhibitory constant (K_i) is shown. **(E)** CDNB assays testing inhibitory properties of haemin, haematin, protoporphyrin IX (PPIX), and myoglobin in a dilution series. **(B, D, E)** Mean and SEM from three independent measurements are shown.

Fig. 5. Assessment of haemin binding to *IrGST1* by haemin-agarose pull down and by VIS-spectrophotometry **(A)** SDS-PAGE of the crude *E. coli* homogenate fractionated by haemin-agarose affinity absorption. Induced homogenate of *E. coli* expressing *IrGST1* was subjected to affinity pull-down by incubating the homogenate with haemin-agarose beads. Red asterisk indicates the protein band eluted with 1 M urea and submitted for mass-fingerprint identification. CDNB activities (nmol CDNB/min/mg protein) of individual fractions are shown below the gel picture, mean $\pm$ SEM from three measurements. Activity of eluted fraction was not determined due to the presence denaturing urea in elution buffer. **(B)** Titration of *IrGST1* was carried out in a range of molar ratios to the constant concentration of 10 μ M haemin. **(C)** A plot of absorbance values at 421 nm in relation to molar ratios of *IrGST1* and haemin.

Fig. 6. Purification and haem-binding of untagged *IrGST1*. **(A)** SDS-PAGE monitoring a two-step purification of untagged *IrGST1* from *E. coli* homogenate (Crude). First step was performed by anion exchange (IEX) chromatography using MonoQ column. Second step was performed by size-exclusion chromatography (SEC) using Superdex 75 column. **(B)** SEC chromatogram of purified untagged *IrGST1* detected at 280 (black line) and 400 nm (red line). The elution volumes of molecular weight standards are indicated by arrows. The deduced molecular weight of *IrGST1* was calculated using a logarithmic standard curve. **(C)** SEC chromatogram detecting both 280 nm (black line) and 400 nm (red lines) of purified *IrGST1* pre-incubated with varying molar ratios of haemin.

Legends to the Supplementary Figures

Supplementary Fig. S1. Multiple amino acid alignment of *IrGST1* sequence with putative homologues of delta-/epsilon-class GSTs from *I. scapularis* genome. *IrGST1* (*Ixodes ricinus*, this work, GenBank MF984398); Right panel table – Nomenclature, VectorBase and GenBank accession Nos of *I. scapularis* genes encoding delta-/epsilon-class GSTs adopted from (Reddy et al., 2011).

Supplementary Fig. S2. Immunohistochemistry evaluation of lipid peroxidation. Sections were prepared from guts dissected from semi-engorged *I. ricinus* females (5 days of feeding). Section were

labelled with rabbit α 4-HNE (1:300) and Alexa488-conjugated anti-rabbit antibody (left). DAPI was used as a counterstain. Sections were also stained with toluidine blue (right) - general structure of the tick gut showing the boundary between the gut epithelium and the gut lumen, containing large haemoglobin crystals (Hb) and digest gut cells (dGC); Nc - nuclei.

Supplementary Fig. S3. Binding of *IrGST1* to glutathione or S-hexyl glutathione agarose. (A) Activity measurements with titrated 1-chloro-2,4-dinitrobenzene (CDNB) concentrations to calculate Michaelis-Menten constant (K_m). (B) Activity measurements with titrated glutathione (GSH) concentrations to calculate Michaelis-Menten constant (K_m). (C) Induced *E. coli* fraction as well as Ni^{2+} -IMAC purified *IrGST1* were subjected to glutathione or S-hexyl glutathione (S6) agarose. Individual fractions were examined by SDS-PAGE and activity measurements. Mean and SEM are shown, $n = 3$. FT - flow-through, E - elution, N.D. - not determined with a given detection limit.

Supplementary Fig. S4. Isoelectric focusing electrophoresis of purified untagged *IrGST1*. Standard pI values of the markers (M) are indicated on the left.

Supplementary Fig. S5. Haem-binding size exclusion chromatography (SEC). SEC chromatogram detecting both 280 and 400 nm of purified untagged *IrGST1* pre-incubated with varying molar ratios of haemin. Arrows point towards a deduced dimeric form of the protein, asterisks denote a higher molecular weight *IrGST1*:haemin complex as a potential polymer.

A)

B)

Highlights

- Haem-inducible *IrGST1* from *Ixodes ricinus* is the first functionally characterised tick GST of delta-/epsilon-class
- *IrGST1* and its orthologues from other ticks form a phylogenetically distinct clade of GSTs that secondarily acquired haem-binding properties
- *ir-gst1* tick gut expression is induced by host haem, but not by host iron
- *IrGST1* binds haemin *in vitro* and is presumably an endogenous intracellular scavenger of excessive haem