Aberystwyth University

An oligonucleotide microarray for transcriptome analysis of Schistosoma mansoni and its application/use to investigate gender-associated gene expression

Fitzpatrick, Jennifer M.; Johnston, David A.; Williams, Gary W.; Williams, Debbie J.; Freeman, Tom C.; Dunne, David W.; Hoffmann, Karl F.

Published in:
Molecular and Biochemical Parasitology

DOI:
[10.1016/j.molbiopara.2005.01.007](https://doi.org/10.1016/j.molbiopara.2005.01.007)

Publication date:
2005

Citation for published version (APA):
Fitzpatrick, J. M., Johnston, D. A., Williams, G. W., Williams, D. J., Freeman, T. C., Dunne, D. W., & Hoffmann, K. F. (2005). An oligonucleotide microarray for transcriptome analysis of Schistosoma mansoni and its application/use to investigate gender-associated gene expression. *Molecular and Biochemical Parasitology*, 141(1), 1-13. <https://doi.org/10.1016/j.molbiopara.2005.01.007>

General rights

Copyright and moral rights for the publications made accessible in the Aberystwyth Research Portal (the Institutional Repository) are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the Aberystwyth Research Portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the Aberystwyth Research Portal

Take down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

tel: +44 1970 62 2400
email: is@aber.ac.uk

An oligonucleotide microarray for transcriptome analysis of *Schistosoma mansoni* and its application/use to investigate gender-associated gene expression[☆]

Jennifer M. Fitzpatrick^a, David A. Johnston^b, Gary W. Williams^c, Debbie J. Williams^c,
Tom C. Freeman^c, David W. Dunne^a, Karl F. Hoffmann^{a,*}

^a Department of Pathology, University of Cambridge, Tennis Court Road, Cambridge CB2 1QP, UK

^b Department of Zoology, The Natural History Museum, Cromwell Road, London SW7 5BD, UK

^c Rosalind Franklin Centre for Genomics Research, Hinxton, Cambridge CB10 1SB, UK

Received 14 September 2004; received in revised form 12 December 2004; accepted 4 January 2005

Abstract

Global profiling transcriptomes of parasitic helminths offers the potential to simultaneously identify co-ordinately expressed genes, novel genetic programs and uniquely utilized metabolic pathways, which together provide an extensive and new resource for vaccine and drug discovery. We have exploited this post-genomic approach to fabricate the first oligonucleotide DNA microarray for gene expression analysis of the parasitic trematode *Schistosoma mansoni*. A total of 17,329 *S. mansoni* DNA sequences were used to design a microarray consisting of 7335 parasite elements or approximately 50% of this parasite's transcriptome. Here, we describe the design of this new microarray resource and its evaluation by extending studies into gender-associated gene expression in adult schistosomes. We demonstrate a high degree of reproducibility in detecting transcriptional differences among biologically replicated experiments and the ability of the microarray to distinguish between the expression of closely related gene family members. Importantly, for issues related to sexual dimorphism, labour division, gamete production and drug target discovery, 197 transcripts demonstrated a gender-biased pattern of gene expression in the adult schistosome, greatly extending the number of sex-associated genes. These data demonstrate the power of this new resource to facilitate a greater understanding into the biological complexities of schistosome development and maturation useful for identifying novel intervention strategies.

© 2005 Published by Elsevier B.V.

Keywords: *Schistosoma*; Helminth; DNA microarray; Gene expression

1. Introduction

Parasite expressed sequence tag (EST) and genomic sequencing projects have proven to be an invaluable resource for parasite gene discovery and have led to the identifica-

tion of numerous putative gene products [1–3]. Despite the large availability of DNA sequence information, true potential of this resource will only be realized upon the assignment of gene function within an actual biological and cellular context, thus leading to the possible functional annotation of many important parasitic genomes. Towards this goal, several investigators have developed DNA microarrays to probe and begin to elucidate the role of specific gene products in the lifestyle, pathogenicity and fundamental biology of multiple parasites [4–6]. This approach, in combination with continued wide ranging genomic and EST sequencing has brought together genomic and functional-genomic data

[☆] Microarray data reported in this paper is available in the ArrayExpress database at EBI under the reference numbers A-MEXP-134 (description of *S. mansoni* oligonucleotide microarray) and E-MEXP-223 (all microarray data).

* Corresponding author. Tel.: +44 1223 333 338; fax: +44 1223 333 741.
E-mail address: kfh24@cam.ac.uk (K.F. Hoffmann).

to reveal new insights into the complex pathogenic parasitic lifestyle.

Gene discovery and functional analysis of schistosomes has also benefited from such an approach [5,7]. These studies have revealed cDNA microarrays to be particularly useful and reliable in identifying gender-associated transcripts in both *Schistosoma mansoni* [5] and *S. japonicum* [7]. However, while sampling small gene subsets proved fruitful for the initiation of these investigations, utilizing a high-throughput approach on a whole genome scale will dramatically increase the transcriptional understanding of parasite sexual biology where current thinking proposes that males and females evolved to maximize independent functional roles important to the survival of the parasite. Presently, the *S. mansoni* genome contains approximately 14,000 predicted genes, and like many other parasites, most of these genes display no database homology and therefore, have no functional annotation [8–10]. Assigning some putative function or association, based on expression profiling by DNA microarray analysis, may provide some of the most promising research areas for elucidating the molecular basis of parasite biology.

Here we describe the design, fabrication and validation of a new DNA microarray for schistosome transcriptome analysis based on the use of long oligonucleotide probes. Long oligonucleotide DNA microarrays were constructed as highly sensitive alternatives to cDNA microarrays due to inherent advantages throughout production and experimental use [11]. The DNA microarray contains 7335 *S. mansoni* probes covering approximately 50% of the total estimated gene complement and was used here to expand upon our previous studies of schistosome conjugal biology. The experiments described provide a dual function: (1) to specifically characterize the fabricated oligonucleotide DNA microarray allowing sensitive, reproducible gene expression results to be generated by multiple users and laboratories; and (2) to further elucidate the expression profiles of male and female adult parasites with the goal of expanding our knowledge relating to sexual maturation, sexual dimorphism, labour division and gamete production. Ultimately, investigations into *S. mansoni* transcriptional mechanisms will likely generate new insights into the development and maintenance of this helminth's dioecious lifestyle, leading to the identification of novel drug targets or vaccine candidates.

2. Materials and methods

2.1. Parasites

Adult male and female *S. mansoni* (NMRI Puerto Rican strain) were perfused from percutaneously infected mice at 7 weeks after challenge with independent batches of 250 cercariae each shed from albino *Biomphalaria glabrata*. After perfusion, both immature and mature worms were counted

and sex-separated. Miracidia used to infect *B. glabrata* were hatched from eggs collected from mouse livers 7 weeks after infection [12].

2.2. Design of *S. mansoni* DNA oligonucleotide probes

S. mansoni DNA elements chosen for oligonucleotide design were selected from EST sequences available from public databases as of June 28, 2002, full-length mRNA and genomic DNA (gDNA) sequences (using NCBI Entrez limits excluding ESTs, STSs, GSSs, TPAs, patents and working drafts) available from public databases as of April 2003 and from one full-length mRNA sequence identified in our laboratory (AY267032—*S. mansoni* arginase). The 16,815 EST sequences were clustered using the CAP3 DNA sequence assembly program [13] into 2076 contigs (representing more than one EST sequence) and 5049 singletons (representing only one EST sequence) for a total of 7125 unique DNA sequence clusters. In addition to these 7125 non-redundant EST clusters, 513 full-length mRNA (some redundancy with respect to EST clusters) and gDNA sequences were included to bring the total number of DNA sequences used as templates for oligonucleotide design to 7638. CAP3 has previously been shown to be tolerant of sequencing errors resulting from single pass sequencing and is effective at differentiating between closely related gene family members [14].

Putative sequence homology of each schistosome DNA element was assigned using the web-based Basic Local Alignment Search Tool (BLASTx) [15] searching against the NCBI protein non-redundant (nr) database. BLASTx hits with an Expect-value (*E*-value) of $\leq 10^{-05}$ were considered significant and the corresponding NCBI protein nr designation was used to annotate the EST contigs and singletons. BLASTx hits with an *E*-value of $\geq 10^{-05}$ were not considered significant and therefore the corresponding schistosome DNA elements obtaining these scores were annotated as 'UNKNOWN'. In a further attempt to annotate these unknown schistosome DNA sequences, they were compared against the *S. mansoni* EST database compiled by Verjovski-Almeida et al. [9] using BLASTn, where DNA elements generating BLASTn bit score values of ≥ 200 were considered a match. Subsequent annotation was provided then by the EST database assigned through additional BLASTx searches of NCBI (using their applied criteria [9]). Sequence similarity assignment was performed for two reasons: (1) to annotate the DNA sequence representations deposited on each DNA microarray; and (2) to identify which DNA strand to select for oligonucleotide design. For BLASTx searches with no significant similarity, the sense strand was selected for oligonucleotide design, unless there was a poly-T tract incorporated at one end of the parent DNA sequence (anti-sense strand was used in this case). Therefore, a small percentage of oligonucleotides deposited on this DNA microarray may have represented the non-coding DNA strand. The labelling procedure used in this study [16] (Klenow incorpo-

ration of Cy-dCTP molecules into double stranded cDNA) fluorescently labels both strands of cDNA, which effectively compensates for this small percentage of oligonucleotides.

All oligonucleotides were 50 bases in length, modified by a 6-C linked 5' amino modification and synthesized at 200 nmol scale (Illumina, SD, CA). Multiple hierarchical tests were performed on each of the 7638 DNA sequences to select the optimum 50-mer oligonucleotide representative for each contig/singleton/mRNA/gene. The European Molecular Biology Open Software Suite (EMBOSS [17]) hosted at SourceForge (<http://emboss.sourceforge.net/>) was used for most of the analyses. Initially, each sequence was required to be a minimum of 50 bases in length. Those sequences passing this first test were then subjected to a series of selection criteria. Using the EMBOSS program 'dan', regions were identified that were within the required GC content of 30–50%. The program 'palindrome' was used to identify regions with no gross secondary structure (defined as having seven or more bases capable of forming a perfect hairpin structure). The program 'freak' was used to identify regions that had no ambiguous nucleotide codes. Additionally, using the program 'RepeatMasker' (<http://www.repeatmasker.org>), regions were identified that contained no low complexity sections, simple repeats or sequences matching the *S. mansoni* SR2 retrotransposon sequence [18]. Using BLASTn, each potential oligonucleotide source region was then compared against all 7638 *S. mansoni* sequences and areas with no matches of 12 or more bases were identified. If this BLAST criterion proved too restrictive to allow regions in a sequence to be used as oligonucleotides, then it was progressively relaxed to ≥ 20 and then ≥ 40 bases, respectively. Starting from the 3' end of each sequence, a maximum of four non-overlapping regions of sequence with a length of 50 bases or more that passed the selections criteria were identified. The T_m of these possible oligonucleotides was determined using the EMBOSS program 'dan', and the one from each sequence with a T_m closest to 72 °C was selected for oligonucleotide design. Originating from the initial 7638 DNA sequences, 7214 passed all of these tests and were used to design 50-mer oligonucleotides. The remaining 424 DNA sequences that failed were again subjected to the same tests, but with the % GC content relaxed to 25–50%. Out of these 424 DNA sequences, 121 could generate a 50-mer oligonucleotide. The remaining 303 sequences failed these tests due to length (<50 bases), high ambiguity or a GC content outside of the required ranges. All sequences were analyzed using the InterProScan package [19] and Gene Ontology (GO) [20] terms were extracted from the output of this analysis using a perl script. A total of 7335 50-mer *S. mansoni* oligonucleotides were designed and, along with control oligonucleotides (*Bacillus subtilis*-specific and *Ara-bidopsis thaliana*-specific DNA elements), represented the oligonucleotide DNA microarray probes used in this study (Table 1). Oligonucleotide DNA microarrays were printed on CodeLink™ Activated Slides (Amine-Binding Slides)

Table 1

S. mansoni oligonucleotide microarray sequence information

Total number of clustered <i>S. mansoni</i> EST sequences	16,815
Number of contigs	2076
Number of singletons	5049
Number of <i>S. mansoni</i> genes (or full-length mRNAs)	513
Number of <i>S. mansoni</i> sequences used for oligonucleotide design	7638
Total number of microarray elements	8160
Number of oligonucleotides designed from <i>S. mansoni</i> sequences	7335
Number of <i>A. thaliana</i> control sequences	120
Number of <i>B. subtilis</i> control sequences	84
Number of buffer/negative control elements	621
Number of sequences displayed in ambiguous direction	1140
Total number of <i>S. mansoni</i> sequences submitted for BLASTx analysis	7335
Sequences displaying no significant similarity	4621
Sequences displaying significant similarity	2714
Sequences displaying significant similarity to hypothetical proteins	3601
Total number of <i>S. mansoni</i> sequences submitted for BLASTn analysis	4621
Sequences displaying no significant similarity	2591
Sequences displaying significant similarity	2030
Sequences displaying significant similarity to hypothetical proteins	1156
Total number of gene ontology (GO) terms assigned	3605
Sequences assigned one or more GO terms	1242
Number of unique GO terms	476
Molecular function	249
Biological process	161
Cell component	66

Summary of elements deposited on microarray reveal the diversity of this new post-genomic resource for investigations into schistosome biology. Complete details relating to derived numbers can be found in Section 2.

(Amersham Biosciences, UK) at the Rosalind Franklin Centre for Genomics Research (Hinxton, UK) at a concentration of 250 ng μl^{-1} .

2.3. Amplified labelled cDNA target synthesis

Cy3/Cy5-dCTP (Amersham Biosciences, UK) labelled cDNA targets were generated through a procedure described by Petalidis et al. [16]. One microgram of *S. mansoni* total RNA was used in a modified mRNA amplification reaction using template-switching PCR (ts-PCR). Optimal PCR cycle number was established empirically by evaluating yield of PCR product with increasing cycle number. Adult female cDNA was labelled with Cy3-dCTP and adult male cDNA labelled with Cy5-dCTP in initial experiments whereas fluorescent labels were reversed for dye-swap experiments.

2.4. Microarray hybridization

Labelled cDNA was re-suspended in hybridization solution (5× SSC, 5× Denhardt's solution, 1 mM sodium pyrophosphate, 50 mM Tris (pH 7.4) and 0.1% SDS) and

denatured at 95 °C for 5 min then at 50 °C for a further 5 min. Microarray hybridization was performed in a humidified chamber at 45 °C for 16–18 h. Three successive, post-hybridization stringency washes were performed ($2 \times \text{SSC}$, $0.1 \times \text{SSC}/0.1\%$ SDS and $0.1 \times \text{SSC}$) for 3-min/wash solution at room temperature with agitation. Image acquisition (16-bit tiff) for the DNA microarray was performed using a GenePix® 4100A (Axon Instruments Inc.) dual channel laser scanner at 10 μm resolution, 100% laser power and PMT levels ranging from 580 to 730. Image analysis used the Microarray Suite extension of IP Lab for Macintosh (Scanalytics) software.

2.5. DNA microarray data filtering

Data analysis was performed as previously described [7]. Briefly, all data was intensity-dependent normalized using the program Vector Xpression 3.1 (Invitrogen® Life Technologies). Expression ratio (Ex. ratio) was thus defined as the normalized- $\text{Log}_2(\text{Cy5}/\text{Cy3})$ ratio. Poor quality spots and low intensity data were filtered and removed by a succession of applied statistical criteria. Initially, the arithmetic mean was calculated for all non-*S. mansoni* spots contained on the microarray (1650 duplicated negative controls, including *B. subtilis*, *A. thaliana* and buffer spots). The mean signal intensity for each hybridized *S. mansoni* element was required to be greater than one standard deviation above the mean of negative controls in at least one channel (Cy5 or Cy3). All data below this value were removed from further analysis. Oligonucleotides passing these filtering criteria had to display Log_2 normalized expression ratios outside of the 90% confidence interval in at least three out of five replicate DNA microarray hybridizations to be included in the final set of differentially expressed, sex-associated transcripts. Correlation coefficients (*R*) for biological replicate hybridizations were derived from a goodness of fit measure of a linear model where values approaching one indicate a high level of agreement. Hierarchical cluster analysis was applied using single linkage and Euclidian distance correlation matrices. Care was taken to ensure all information is MIAME [21] compliant and all data was submitted to MIAMExpress at European Bioinformatics Institute, Hinxton, UK (<http://www.ebi.ac.uk/miamexpress/>) [22].

2.6. RT-PCR analysis

Reverse transcription PCR was carried out using 1 μg of parasite total RNA as described [23]. All RNA was treated with DNase I (Ambion Inc.) prior to reverse transcription to remove any potential genomic DNA contamination. The oligonucleotide primers for PCR are shown in Table 2 (see supplementary information). All amplicons were separated on a 2% agarose gel, detected using ethidium bromide and images captured using digital photography.

3. Results

3.1. *S. mansoni* sequence analysis and fabrication of a novel oligonucleotide microarray to profile schistosome gene expression

Clustering of the parasite EST sequences contained in NCBI protein nr databases as of June 2002 (using the CAP3 program) and inclusion of full-length genes as of April 2003, led to a collection of 7638 sequences available for oligonucleotide design. Selective criteria for design of optimal oligonucleotides enabled 7335 *S. mansoni* oligonucleotides (duplicated on microarray to 14,670) to be synthesized and arrayed (Table 1). BLASTx manual annotation of each contig/EST revealed 4621 sequences displaying no significant similarity using BLASTx *p*-value (*E*) criteria of $\leq 10^{-05}$, representing $\sim 63\%$ of the total *S. mansoni* sequences. Therefore, from the initial BLASTx annotation, 2714 sequences displayed significant similarity to known genes in NCBI ($\sim 37\%$ of total *S. mansoni* sequences). The 4621 sequences without similarity were then submitted to BLASTn analysis at the University of Sao Paulo/ONCA *S. mansoni* EST database [9,24] and hits with a bit score value of ≥ 200 were merged with the sequences to create longer contigs. These new contigs were then re-submitted to the NCBI nr protein database using BLASTx in an attempt to annotate them. This left 2591 sequences still with no significant similarity to entries deposited in NCBI (using same BLASTx criteria as previously described). In contrast, 2030 sequences gained some annotation through this second database search, 1156 of which having significant similarity to hypothetical proteins (874 of these containing some functional annotation). Therefore, the total number of *S. mansoni* sequences used for oligonucleotide design displaying some annotation is approximately 65% (49% without hypothetical proteins). Additionally, 1140 sequences were potentially presented in the databases in the anti-sense direction and therefore the final oligonucleotide was designed from the sense strand (see Table 1).

All sequences were further analyzed for putative function using the InterProScan package [19] and GO terms were assigned to those sequences with recognized potential functional domains. Briefly, 3605 GO terms were assigned to 1242 *S. mansoni* sequences (Table 1). The GO terms assigned (476 unique) included 249 molecular function, 161 biological process and 66 cell component terms. This reveals the extensive range of potential biological activities and processes represented by the *S. mansoni* sequences deposited on this DNA microarray (see supplementary information).

3.2. Adult gender-associated transcripts are reproducibly detected by fabricated *S. mansoni* oligonucleotide microarrays

As multi-gene families are represented throughout the *S. mansoni* genome (cathepsins, superoxide dismutase, dynein

light chains etc.) it was important to ensure that the 50-mer representation of each DNA element was capable of distinguishing among individual members. Results presented here suggest that multiple *S. mansoni* oligonucleotides representing various members of closely related gene families are able to distinguish among gene family members (Fig. 1). Expression values show that the oligonucleotides were designed with optimal physical properties to allow sensitive and reproducible hybridization, e.g., consistent T_m , minimal secondary structure, minimal sequence similarity to other oligonucleotides within the array (not representing the same gene), not designed over repeat regions or other regions of low complexity. Utilizing two previously reported and well-characterized superoxide dismutase (SOD) genes as a model [25,26], expression values were reported for multiple oligonucleotides representing these genes. Each independent oligonucleotide representing different regions of the same gene, gave comparable expression values as illustrated in Fig. 1A and B. Five oligonucleotides representing exon regions of extracellular superoxide dismutase (M27529) all demonstrated female-associated gene expression values (in agreement with results obtained by Fitzpatrick et al. [7] using a *S. japonicum* cDNA microarray). Normalized $\log_2$ expression values ranged from -3.4 to -6.1 as means of five separate experiments. This finding was independently confirmed by gene-specific RT-PCR in comparison to the constitutively expressed (in adult parasites [27]) α -tubulin gene. In contrast, four oligonucleotides representing cytosolic superoxide dismutase (M97298) (Fig. 1B) showed equal expression between the two sexes (again in agreement with Fitzpatrick et al. [7]). Furthermore, oligonucleotide (a) (Fig. 1B) displays zero expression values (no signal in either channel above background—NS), being representative of an intron region of the cytosolic SOD gene. Additional confirmation by RT-PCR demonstrated the equivalent expression of cytosolic SOD between male and female *S. mansoni* parasites.

S. mansoni oligonucleotide microarrays performed with a high degree of precision and were reproducible in generating similar expression results from five independent biologically replicated experiments. Each individual microarray experiment represented a biological replication of an adult male and adult female comparison (i.e. different batches of worms removed from mice infected with different independent batches of cercariae). Analyzing the reproducibility between these individual and discrete experiments suggest that the microarrays are capable of generating information which is directly comparable between multiple and distinct biological batches of parasites. Scatterplots comparing the expression values from one biological experiment to a different biological experiment demonstrated a high correlation coefficient between experiments (Fig. 1C), these representations displaying $R=0.848$ (experiments 3 and 5, $n=4524$) and $R=0.816$ (experiments 2 and 5, $n=4645$), respectively. The comparisons were performed using all oligonucleotides that passed the initial filtering criteria

(expression values above background levels in the experiments compared, as calculated from negative controls, see Section 2).

The number of genes detected as expressed in the adult ranged from 4678 to 5169 (mean 4973). This figure equates to around 65% of the oligonucleotides deposited on the microarray. Taking into account some redundancy within the arrayed DNA elements, this percentage is comparable with that predicted by Verjovski-Almeida et al. [9] where it was estimated by two independent methods, that $\sim 50\%$ of the gene complement ($\sim 14,000$ genes) of *S. mansoni* was expressed in the adult stage.

3.3. Expression profiling of sexually mature adult-stage *S. mansoni* using the oligonucleotide DNA microarray reveals novel gender-associated transcripts

As quality control experiments illustrated the reproducible nature of this *S. mansoni* DNA oligonucleotide microarray, analyzing the differences in gene expression between adult female and adult male parasites initially validated this resource. Here, all transcripts identified as differentially expressed displayed $\log_2$ normalized gene expression ratios outside of the 90% confidence interval in three out of five independent, biologically replicated experiments. A hierarchical clustergram (single linkage analysis/Euclidian distance correlation) shown in Fig. 2 displays all transcripts identified as being highly differentially expressed in either male (red: positive ratio) or female (green: negative ratio) parasites. These male/female bimodal comparisons revealed 141 unique genes highly expressed in the female when compared to the male, and 86 genes highly expressed in the male when compared to the female. This considerably increases the number of known sex-associated transcripts in *S. mansoni* adult parasites. Importantly, dye-swap experiments showed a high correlation with data passing these strict filtering criteria and therefore suggest minimal dye effect on the prediction of differentially expressed transcripts. Twenty-four oligonucleotides (16 classified as having no significantly sequence similarity) highly expressed in adult females did not reproduce as differentially expressed when compared to adult male in multiple pooled RNA dye-swap experiments. Moreover, only six oligonucleotides described as male associated did not repeat using the dye-swap experiments. Therefore, after considering the minimal dye effect observed in our experiments, 117 unique genes were highly expressed in the adult female and 80 unique genes were highly expressed in the adult male (see supplementary information for full list of oligonucleotides identified as differentially expressed between the adult genders).

Gene expression results for all differentially expressed transcripts relate to the vast majority of previously appreciated gender-associated genes. Microarray expression values reveal these putative positive controls to be differentially expressed in the previously documented gender-associated

Fig. 1. *S. mansoni* oligonucleotide DNA microarrays are capable of generating reproducible expression results and can distinguish transcriptional differences among gene family members. (A) Hybridization performance of multiple oligonucleotides spanning different intragenic regions of two superoxide dismutase homologues. Five oligonucleotides spanning the three extracellular superoxide dismutase exons (Ex-SOD, M27529) reveal similar gene expression profiles (for all five biological replicates averaged) and strongly support the female-enriched expression of this SOD homologue (verified by gene-specific RT-PCR). (B) Cytosolic SOD (Cyt-SOD, M97298) did not display a statistically significant gender association as determined from hybridization information originating from four oligonucleotides spanning two exons and the 3'-UTR. No signal intensity (NS) was observed for a Cyt-SOD specific oligonucleotide designed over an intron. Boxes represent exons, solid lines represent non-coding DNA elements and dashed lines represent positions of designed oligonucleotides for the 5.1 kb Ex-SOD and the 8.5 kb Cyt-SOD genes. RT-PCR conditions and derivation of expression ratio (Ex. ratio) are described in Section 2. (C) Statistical analysis of biologically reproduced DNA microarray hybridization experiments. Scatterplots compare the Log₂-calibrated ratio generated for each oligonucleotide probe (mean of two replicate spots) from one biological batch of adult worm material to the Log₂-calibrated ratio generated for the same oligonucleotide probe (mean of two replicate spots) from a different biological batch. The correlation coefficient values from two representative comparisons, $R = 0.848$ and 0.816 indicate a high degree of agreement between biologically replicated experiments. Scatterplots display oligonucleotide probes that passed the initial filtering criteria (signal intensity greater than 1 S.D. above the mean of negative control elements) for quality data in each biological batch/hybridization comparison and include 4524 for experiments 3 and 5, and 4645 elements for experiments 1 and 5. Lines represent the line of regression (centre line) and the predicted 99% confidence intervals of the plotted data.

Fig. 2. *S. mansoni* oligonucleotide microarrays can reproducibly detect gender-enriched gene expression profiles in sexually mature *S. mansoni* male and female adult parasites. Single-linkage hierarchical clustering (using Euclidean distance correlation matrices) of all gender-enriched transcripts passing filtering criteria from five independent experimental replicates and dye-swap experiments. Representative gender-enriched transcripts verified by RT-PCR (Fig. 3) are listed next to the clustergram along with their unique identifier (contig ID, accession number or name). All other gender-enriched transcripts identified in this study can be found in accompanying [supplementary information](#). Increasing shades of red depict greater expression in adult male parasites, increasing shades of green represent greater expression in adult female parasites and grey represents gene expression data absent for that particular replicate experiment.

manner. These transcripts include those previously identified in our laboratory [5,7] as well as female eggshell proteins (p48/chorion/34 kDa) [28–33], female-specific 800 protein [34], ferritin-1 [35], ORF-RF2 [36], glutathione peroxidase [37] and adenylosuccinate lyase [38] amongst others. These positive controls, confirming predicted patterns of gene expression, again reinforced the usefulness of this oligonucleotide microarray to reveal new and interesting gender-associations. A large number of novel associations were also observed, revealing both adult female and adult male expression biases. These associations included the female-enriched expression of an extracellular superoxide dismutase and tyrosinase 1 and 2, although all of these have previously been reported as female-associated in *S. japonicum* [7] (and *S. mansoni* for tyrosinase) [5]. Further novel female-associated transcripts included an acyl-CoA diacylglycerol acyltransferase (DGAT), acyl-CoA cholesterol acyltrans-

ferase (ACAT), UDP-GlcNAc:Alpha-6-D-Mannoside Beta-1, 2-N-Acetylglucosaminyltransferase II, multiple histidine-rich proteins, serine-threonine proteins, a large amino acid transporter (also previously observed in *S. japonicum* [7]), anti-inflammatory protein-16, stathmin-like protein (SPL) and purine-nucleoside phosphorylase. Furthermore, a significant proportion of all oligonucleotides identified as differentially expressed in the female showed no significant identity with sequences contained in the NCBI protein nr databases. Analysis revealed that this list of female-associated transcripts contained 39 such sequences and an additional 25 sequences annotated as putative hypothetical proteins.

An extensive list of male-associated transcripts was also established; some confirm previous experiments [5,7] but most report novel correlations. Since the number of known male-associated genes has been limited, these data have

brought a variety of new information available in this area of schistosome biology. Muscle, components of the tegument and cytoskeletal elements dominate this set of male-associated transcripts, and include myosin (multiple oligonucleotides representing multiple subunit chains), paramyosin, tropomyosin, alpha-actinin, annexin, fimbrin, microtubule-associated protein 1B, echinonectin, Sm8, troponin T, troponin I and Sm20. Other interesting male-associated transcripts include those inferred to be a ryanodine receptor, procollagen-lysine 2-oxoglutarate 5-dioxygenase 1 precursor (lysyl hydroxylase 1), two collagens, an extracellular superoxide dismutase (contig1384; a different isoform to that detected as female-expressed) and the high voltage-activated calcium channel beta subunit Cav_B1, believed to be partially responsible for praziquantel sensitivity [39,40] (although this transcript did not repeat in the dye-swap experiment, subsequent RT-PCR analysis (Fig. 3B) suggests a definite male bias for its expression). Additionally, 16 sequences with no significant similarity to genes in NCBI and 24 hypothetical proteins were also identified.

See [supplementary information](#) for full list of differentially expressed transcripts for both female and male adult parasites.

3.4. Verification of gender-biased transcripts by RT-PCR

Confirmation of microarray expression data by an independent method of analysis demonstrated the precision of the transcriptional-profiling facilitated by this new oligonucleotide DNA microarray resource. Gene-specific primers were used in an RT-PCR reaction to confirm the expression profiles detected by microarray analysis. RT-PCR results showed the exact same pattern of predicted gene expression revealed by microarray hybridization. Sixteen randomly selected female-associated (by DNA microarray analysis) transcripts were tested and shown to be strongly female expressed by RT-PCR in comparison to the male (using the α -tubulin gene as an internal standard and sample control [27] Fig. 3A). In addition, the 16 male transcripts chosen for confirmation were also identified as more heavily expressed in the male when compared to the adult female by RT-PCR analysis (Fig. 3B). One microarray oligonucleotide representing glutathione peroxidase (SCMGPX1A, L37762) was designed over an intronic region of the gene (and therefore displayed no expression values). As predicted by previous studies [37] and shown here by RT-PCR, this transcript was female-associated. All transcripts chosen (randomly) for RT-PCR verification showed the same pattern of expression as in the DNA microarray analysis, thus demonstrating the usefulness of both the microarray resource and the filtering criteria applied. Transcripts denoted with ‡ did not reproduce male-associated expression in dye-swap experiments (did not pass strict filtering criteria), however, were revealed as highly expressed in the male when the original dye combination was used.

4. Discussion

DNA microarray analysis is now a well-established functional genomics tool for the global analysis of gene expression [41,42]. The oligonucleotide microarray fabricated here represents ~50% of the estimated *S. mansoni* gene complement [9]. The sequences utilized for oligonucleotide design are proportionally representative of the entire *S. mansoni* genome, since the relative percentages of sequences displaying significant similarity to known genes in the databases is similar to that estimated by large-scale EST sequencing efforts [9]. Furthermore, the distribution of sequences between distinct biological functions and processes reflects the transcriptome of this parasite as a whole and suggests diverse roles in numerous biological functions for independent sequences. Since the sequences examined here represent a range of important processes and originate from diverse developmental stages, this established DNA microarray has major implications for probing different biological questions.

Although DNA microarray technology is relatively widely used, this is the first time long DNA oligonucleotides have been employed for large-scale profiling of gene expression in *S. mansoni*. Consequently, in order to characterize, optimize and assess this post-genomics tool, a set of experiments profiling the transcriptome of mature adult schistosomes was performed, similar to previous studies [5,7]. While sexually mature, adult male and female parasites transcribe the majority of genes to a similar degree, it is anticipated that numerous genes will be differentially expressed, as the sexes are morphologically, functionally and chromosomally distinct. Having evolved from hermaphroditic ancestors, it is likely that differential gene expression has driven sexual dimorphism and labour division within the genus and ultimately led to cooperative conjugal biology as a highly successful means of maximizing parasite transmission. Although highly effective for continuing the lifecycle, intravascular conjugal biology resulting in egg production by adult female parasites also contributes to host-mediated, inflammatory, circumoval immune responses. If uncontrolled, these immune responses can precipitate a series of pathological complications resulting in severe morbidity and mortality in infected individuals [43]. Since the transcriptional basis of adult sexual maturation and egg-production remains relatively unknown, any information gained will be crucial to the elucidation of the specific processes involved. To be classified here as differentially expressed between the sexes, genes had to pass independent statistical criteria: (1) expression values had to be significantly above background levels in order to remain in the dataset (as calculated from negative control elements); and (2) genes had to be outside of the 90% confidence interval of the entire range of expressed data in three of five independent biologically replicated hybridizations and passed the identical criteria in multiple dye-swap experiments using pooled RNA samples from independent batches. These criteria enabled a total of 117 female and 80 male genes to be reproducibly and confidently identified as differentially ex-

Unique ID	M	F	NCBI Similarity (BLASTx)	PCR cycle #
Contig1301			Diacylglycerol Acyltransferase 2-like (3E ⁻⁵⁴)	40
Contig1536			CD36 Antigen (LAMP-3) (5E ⁻¹⁵)	29
SCMGPX1A (*)			Glutathione peroxidase (0.0)	29
Contig2828			CD36 Antigen-like 2 (LIMP-II) (3E ⁻²³)	29
AF269252			Anti-inflammatory protein Sm16	40
Contig1085			Polo-like kinase homolog (5E ⁻⁷⁶)	40
AF091509			Strathmin-like protein (0.0)	40
Contig1636			Cytoplasmic aspartate aminotransferase (2E ⁻⁶⁴)	40
SCMSODM			Extracellular superoxide dismutase (0.0)	40
Contig4115			Vesicular integral membrane protein Vip36 (7E ⁻³⁷)	40
Contig1015			Sterol O - acetyltransferase 1 (ACAT) (5E ⁻²⁷)	40
Contig5379			Large neutral amino acid transporter small subunit 2 (4E ⁻²⁹)	40
Contig844			Limulus factor D (4E ⁻¹⁰)	40
SCMGLUPER			Glutathione peroxidase (0.0)	35
Contig948			SmTYR1 (tyrosinase 1) (0.0)	26
Contig1617			SmTYR2 (tyrosinase 2) (0.0)	35
Contig1475 (A)			UDP-GLCNAC:Alpha-6-D-mannoside beta-1, 2-N-acetylglucosaminyltransferase II (1E ⁻³⁹)	32
Contig4516			Echinonectin (5E ⁻²³)	35
Contig4386			Extensin class I (3E ⁻⁰⁶)	35
Contig5545			Major tegumental antigen Sm15 (1E ⁻³⁹)	28
Contig1384			Extracellular superoxide dismutase precursor (1E ⁻²³)	40
SCMCALPAIN			Calpain (0.0)	29
AY033598.1 ‡			High voltage-activated calcium channel beta subunit CavB1	29
Contig4341			LIM-domain protein LMP-1 (4E ⁻⁰⁵)	29
SMU19945			Actin 2 (0.0)	22
Contig1777			Desmoyokin (1E ⁻⁵⁰)	28
Contig5696			Procollagen-lysine, 2-oxoglutarate 5-dioxygenase 1 (5E ⁻²⁹)	40
Contig6802			Collagen alpha 1 (XI) chain (3E ⁻¹⁴)	35
Contig5158			Low density lipoprotein-receptor related protein (2E ⁻¹⁰)	35
SCMGTPA1			Glucose transporter protein-1 (0.0)	35
Contig2712 ‡			Nidogen 1 (7E ⁻⁰⁶)	29
Contig5526			Fibrillar collagen chain FAp1 alpha (1.1E ⁻⁷³)	40
AF313910			Tegumental antigen (Sm8) mRNA, partial cds (0.0)	33
(B) M80214			α -tubulin (0.0)	26

Fig. 3. RT-PCR analysis confirmed gender-enriched DNA microarray data for all transcripts tested, illustrating a high correlation between global gene expression profiling and individual gene transcript abundance. One microgram of pooled (five experimental replicates) *S. mansoni* total RNA (DNase I treated (Ambion Inc.)) was used in a RT reaction to prime cDNA synthesis as described in Section 2. Unique identifier (Unique ID) and BLASTx NCBI nr protein database annotation of amplified transcript, sequence of each PCR primer pair, cycle number, expected product size and annealing temperatures for each verified cDNA are listed in Table 2, [supplementary information](#). (A) RT-PCR confirmation of randomly selected female-enriched transcripts identified by DNA microarray analysis. (B) RT-PCR confirmation of randomly selected male-enriched transcripts identified by DNA microarray analysis. Amplification of constitutive α -tubulin served as an internal control for all PCR reactions. (*) indicates female-enriched expression of a glutathione peroxidase ortholog determined solely by RT-PCR as oligonucleotides for SCMGPX1 deposited on DNA microarray were designed over intronic regions. (‡) denotes two transcripts that did not pass filtering criteria for differential expression in the dye-swap experiments. BLASTx E-values for amplified gene products are given to indicate sequence similarity.

pressed, revealing a massive amount of new molecular data. Many positive controls confirmed the validity of these results, including the female-associated eggshell proteins, as previously demonstrated in *S. mansoni* [5] and *S. japonicum* cDNA [7] microarray experiments.

Investigating the putative function of many of these gender-associated transcripts has revealed a number of interesting and previously unexplored patterns of expression. It is now clear that the vast majority of male-associated transcripts are intimately involved in the structural organization of the parasite (supplementary information) which contrasts directly with those transcripts highly expressed in the adult female, some having a presumed role in reproductive and egg-laying mechanisms. Structural elements expressed differentially by the male include components of the muscular system, tegumental proteins and the underlying cytoskeleton. This apparent 'division of labour' between the genders [44,45] now has an extensive transcriptional basis. The male ensures the survival of the egg-laying female by providing physical support and musculature to aid feeding [46], physical transportation within the vasculature [47] and potential extracellular maturation factors [48,49]. The female is thus allowed to concentrate energy expenditure on egg-production, as reflected by the large number of transcripts involved in reproduction and development.

Differentially expressed transcripts of special interest were revealed in both genders (a number of which were subsequently confirmed by RT-PCR analysis, Fig. 3). Evidence exists to the importance of regulated intracellular calcium levels to parasitism by *S. mansoni*; Ca^{2+} signalling is absolutely required for muscular contraction (important more so in the male given their extensive muscular scaffold and infrastructure). Multiple voltage-gated Ca^{2+} channels have been identified within schistosomes, including both α - and β -subunits of *S. mansoni* (and *S. japonicum*) [50,51]. Here, microarray analysis showed the expression of SmCaV_B1 to be significantly greater in the male than the female (in direct contrast to SmCaV_B2 , which appears to be similarly expressed between the two genders). Although in one dye combination, SmCaV_B1 did not demonstrate a significantly strong male association, RT-PCR analysis of this transcript (Fig. 3) confirmed the original microarray results and suggested that dye-swap experiments can be influenced at multiple levels and should be interpreted with caution. Praziquantel (PZQ), the current drug of choice for the treatment of schistosomiasis, is believed to instigate disruption of Ca^{2+} homeostasis within the parasite, although the exact mechanism is unknown [52,53]. It has become clear in recent years that female parasites are much less sensitive to the action of PZQ than males [54], the SmCaV_B1 subunit being intimately involved in the conference of sensitivity [39]. Moreover, these DNA microarray results also revealed a male-associated expression of desmoyokin (AHNAK protein [55]). Although AHNAK may possess multiple functions within a cell, it has been shown to interact specifically with the β -subunits (1, 2 and 2a) of the L-type Ca^{2+} channel and with *F*-actin of hu-

man heart muscle, mediating directly Ca^{2+} signal transduction [56]. It is possible then that high levels of SmCaV_B1 and AHNAK in the male may be instrumental in their increased sensitivity to PZQ observed when compared to females. Numerous other important Ca^{2+} -binding proteins (CaBP) were additionally identified as being male-associated, including calpain (calcium-activated neutral protease (CANP)) [57], calmodulin [58], Sm20 [59] and a ryanodine receptor [60].

From the list of differentially expressed transcripts, females express a higher proportion of enzymes than do males. Included here as specifically female-associated is the expression of two endoplasmic reticulum neutral lipid synthesis enzymes acyl-CoA:diacylglycerol acyltransferase 2-like (DGAT2-like) and acyl-CoA:cholesterol-acyltransferase-1 (ACAT1). ACAT covalently joins cholesterol and fatty acyl-CoA molecules to form cholesterol esters [61–63] and DGAT catalyzes a similar reaction to generate triglycerides, using diacylglycerol as the acyl group acceptor [64,65]. Subsequent sequence analysis and database queries identified a further two potential DGAT1 and DGAT2-like molecules in *S. mansoni*. RT-PCR analysis (in comparison to α -tubulin) revealed a female-associated expression bias for each of these four genes (data not shown). *S. mansoni* is known to be unable to synthesize fatty acids and sterols de novo [66]. The specific role of these enzymes in the female at this time is still unclear, although the generation of triglyceride stores within the parasite is likely to represent a major function. Since schistosomes do not generate energy/ATP through the β -oxidation of fatty acids [67] (despite much of the enzymatic capacity for this being encoded within the genome, data not shown), the parasites may use neutral lipids as stores to guard against high intracellular (toxic) levels of free fatty acids and cholesterol. In addition, the adult female may utilize neutral lipids during egg production.

Concomitant with the egg-laying process, the female consumes a large excess of red blood cells in comparison to the male [68]. The female therefore requires the by-products of haemoglobin digestion to be metabolized, eliminated and stored to reduce direct toxicity and lipid peroxidation. The high expression of the enzymes superoxide dismutase and aspartate aminotransferase, together with the previously known female-associated cathepsins and aspartic proteases [69] and ferritin-1 (stores Fe^{3+} in non-toxic form) [35] in the female, as shown by both DNA microarray analysis and RT-PCR, suggests these enzymes may share a functional role in this process.

The DNA microarray analysis described in this study and combined with previous studies [5,7] revealed a far greater number of transcripts displaying female-associated expression than male. This may be directly indicative of the enhanced metabolic requirements of extensive egg-production, but in addition, may also reflect the presence of actual egg transcripts within the female sample pool. Current investigations utilizing this oligonucleotide microarray to longitudinally analyze the development of the parasite within the host will identify such transcripts and enable a more detailed

analysis of the transition between female and egg as well as subsequent maturation throughout the lifecycle. It is important and timely to note here that numerous transcripts were additionally analyzed at the protein/enzyme activity level. All enzyme activities tested, including tyrosinase and aspartate aminotransferase correlated directly with the predicted pattern of mRNA transcript expression (data not shown).

This study suggests that the design and use of an oligonucleotide microarray to profile the *S. mansoni* transcriptome, is both a feasible and an efficient strategy to examine transcriptional differences between parasite life-stages, time-points, drug challenges and physiological conditions, for example. This oligonucleotide DNA microarray is presently being employed by other laboratories interested in different aspects of schistosome biology and will facilitate joint and collaborative efforts towards a better understanding of this important pathogen. Future investigations into individual functions, specifics and potential interactions of these newly-defined and potential fundamentally-important genes will further define their differential role(s) in the conjugal biology of schistosomes (several of these functional studies are currently ongoing). Highlighting the potential functional properties of unknown genes, by virtue of their expression profiles, has provided priorities and avenues for further research. Whole genome sequence annotation, in conjunction with further DNA microarray analyses, along with other techniques, such as signal-sequence trap [70], RNAi [71,72], proteomics [73] and investigations of large-scale protein–protein interactions [74] provides prospects to dramatically improve the current understanding of the molecular basis of pathogenicity and pathology by this complex and important major parasite of humans.

Acknowledgements

This work was supported by means of a Wellcome Trust Career Development grant awarded to KFH and a UNDP/WORLD BANK/WHO Special Programme for Research and Training in Tropical Diseases (TDR) grant awarded to DAJ. We thank Maureen Laidlaw and Frances Jones for excellent technical help with *S. mansoni* lifecycle maintenance, Dr. Patrick Emery for assistance with EST clustering and Andrew McArdle for computer support.

Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.molbiopara.2005.01.007.

References

- [1] Santos TM, Johnston DA, Azevedo V, et al. Analysis of the gene expression profile of *Schistosoma mansoni* cercariae using the expressed sequence tag approach. *Mol Biochem Parasitol* 1999;103(1):79–97.
- [2] Bahl A, Brunk B, Crabtree J, et al. PlasmoDB: the *Plasmodium* genome resource. A database integrating experimental and computational data. *Nucleic Acids Res* 2003;31(1):212–5.
- [3] Merrick JM, Osman A, Tsai J, et al. The *Schistosoma mansoni* gene index: gene discovery and biology by reconstruction and analysis of expressed gene sequences. *J Parasitol* 2003;89(2):261–9.
- [4] Hayward RE, Derisi JL, Alifadhli S, et al. Shotgun DNA microarrays and stage-specific gene expression in *Schistosoma mansoni* malaria. *Mol Microbiol* 2000;35(1):6–14.
- [5] Hoffmann KF, Johnston DA, Dunne DW. Identification of *Schistosoma mansoni* gender-associated gene transcripts by cDNA microarray profiling. *Genome Biol* 2002;3(8):RESEARCH0041.
- [6] Singh U, Brewer JL, Boothroyd JC. Genetic analysis of tachyzoite to bradyzoite differentiation mutants in *Schistosoma mansoni* reveals a hierarchy of gene induction. *Mol Microbiol* 2002;44(3):721–33.
- [7] Fitzpatrick JM, Johansen MV, Johnston DA, Dunne DW, Hoffmann KF. Gender-associated gene expression in two related strains of *Schistosoma mansoni*. *Mol Biochem Parasitol* 2004;136(2):191–209.
- [8] Gardner MJ, Hall N, Fung E, et al. Genome sequence of the human malaria parasite *Schistosoma mansoni*. *Nature* 2002;419(6906):498–511.
- [9] Verjovski-Almeida S, DeMarco R, Martins EA, et al. Transcriptome analysis of the acelomate human parasite *Schistosoma mansoni*. *Nat Genet* 2003;35(2):148–57.
- [10] Hu W, Yan Q, Shen DK, et al. Evolutionary and biomedical implications of a *Schistosoma mansoni* complementary DNA resource. *Nat Genet* 2003;35(2):139–47.
- [11] Hughes TR, Mao M, Jones AR, et al. Expression profiling using microarrays fabricated by an ink-jet oligonucleotide synthesizer. *Nat Biotechnol* 2001;19(4):342–7.
- [12] Smithers SR, Terry RJ. The infection of laboratory hosts with cercariae of *Schistosoma mansoni* and the recovery of the adult worms. *Parasitology* 1965;55(4):695–700.
- [13] Huang X, Madan A. CAP3: a DNA sequence assembly program. *Genome Res* 1999;9(9):868–77.
- [14] Liang F, Holt I, Pertea G, et al. An optimized protocol for analysis of EST sequences. *Nucleic Acids Res* 2000;28(18):3657–65.
- [15] Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. Basic local alignment search tool. *J Mol Biol* 1990;215(3):403–10.
- [16] Petalidis L, Bhattacharyya S, Morris GA, et al. Global amplification of mRNA by template-switching PCR: linearity and application to microarray analysis. *Nucleic Acids Res* 2003;31(22):e142.
- [17] Rice P, Longden I, Bleasby A. EMBOS: the European molecular biology open software suite. *Trends Genet* 2000;16(6):276–7.
- [18] Drew AC, Minchella DJ, King LT, Rollinson D, Brindley PJ. SR2 elements, non-long terminal repeat retrotransposons of the RTE-1 lineage from the human blood fluke *Schistosoma mansoni*. *Mol Biol Evol* 1999;16(9):1256–69.
- [19] Zdobnov EM, Apweiler R. InterProScan—an integration platform for the signature-recognition methods in InterPro. *Bioinformatics* 2001;17(9):847–8.
- [20] Gene Ontology Consortium (<http://www.geneontology.org/>).
- [21] MIAME (<http://www.mged.org/Workgroups/MIAME/miame.html>).
- [22] MIAMExpress (<http://www.ebi.ac.uk/miamexpress/>).
- [23] Wynn TA, Eltoum I, Cheever AW, et al. Analysis of cytokine mRNA expression during primary granuloma formation induced by eggs of *Schistosoma mansoni*. *J Immunol* 1993;151:1430–40.
- [24] *Schistosoma mansoni* EST Project. OSmEG (<http://verjo18.iq.usp.br/schisto/>).
- [25] Hong Z, Kosman DJ, Thakur A, Rekosh D, LoVerde PT. Identification and purification of a second form of Cu/Zn superoxide dismutase from *Schistosoma mansoni*. *Infect Immun* 1992;60(9):3641–51.
- [26] Simurda MC, van Keulen H, Rekosh DM, LoVerde PT. *Schistosoma mansoni*: identification and analysis of an mRNA and a gene encoding superoxide dismutase (Cu/Zn). *Exp Parasitol* 1988;67(1):73–84.

- [27] Webster PJ, Seta KA, Chung SC, Mansour TE. A cDNA encoding an alpha-tubulin from *Schistosoma mansoni*. *Mol Biochem Parasitol* 1992;51(1):169–70.
- [28] Bobek LA, Rekosh DM, LoVerde PT. Small gene family encoding an eggshell (chorion) protein of the human parasite *Schistosoma mansoni*. *Mol Cell Biol* 1988;8(8):3008–16.
- [29] Chen LL, Rekosh DM, LoVerde PT. *Schistosoma mansoni* p48 eggshell protein gene: characterization, developmentally regulated expression and comparison to the p14 eggshell protein gene. *Mol Biochem Parasitol* 1992;52(1):39–52.
- [30] Henkle KJ, Cook GA, Foster LA, et al. The gene family encoding eggshell proteins of *Schistosoma mansoni*. *Mol Biochem Parasitol* 1990;42(1):69–82.
- [31] Johnson KS, Taylor DW, Cordingley JS. Possible eggshell protein gene from *Schistosoma mansoni*. *Mol Biochem Parasitol* 1987;22(1):89–100.
- [32] Koster B, Dargatz H, Schroder J, et al. Identification and localisation of the products of a putative eggshell precursor gene in the vitellarium of *Schistosoma mansoni*. *Mol Biochem Parasitol* 1988;31(2):183–98.
- [33] Rodrigues V, Chaudhri M, Knight M, et al. Predicted structure of a major *Schistosoma mansoni* eggshell protein. *Mol Biochem Parasitol* 1989;32(1):7–13.
- [34] Reis MG, Kuhns J, Blanton R, Davis AH. Localization and pattern of expression of a female specific mRNA in *Schistosoma mansoni*. *Mol Biochem Parasitol* 1989;32(2–3):113–9.
- [35] Dietzel J, Hirzmann J, Preis D, Symmons P, Kunz W. Ferritins of *Schistosoma mansoni*: sequence comparison and expression in female and male worms. *Mol Biochem Parasitol* 1992;50(2):245–54.
- [36] Bobek L, Rekosh DM, van Keulen H, LoVerde PT. Characterization of a female-specific cDNA derived from a developmentally regulated mRNA in the human blood fluke *Schistosoma mansoni*. *Proc Natl Acad Sci USA* 1986;83(15):5544–8.
- [37] Roche C, Liu JL, LePresle T, Capron A, Pierce RJ. Tissue localization and stage-specific expression of the phospholipid hydroperoxide glutathione peroxidase of *Schistosoma mansoni*. *Mol Biochem Parasitol* 1996;75(2):187–95.
- [38] Foulk BW, Pappas G, Hirai Y, Hirai H, Williams DL. Adenylsuccinate lyase of *Schistosoma mansoni*: gene structure, mRNA expression, and analysis of the predicted peptide structure of a potential chemotherapeutic target. *Int J Parasitol* 2002;32(12):1487–95.
- [39] Kohn AB, Roberts-Misterly JM, Anderson PA, Khan N, Greenberg RM. Specific sites in the beta interaction domain of a schistosome Ca^{2+} channel beta subunit are key to its role in sensitivity to the anti-schistosomal drug praziquantel. *Parasitology* 2003;127(Part 4):349–56.
- [40] Kohn AB, Roberts-Misterly JM, Anderson PA, Greenberg RM. Creation by mutagenesis of a mammalian Ca^{2+} channel beta subunit that confers praziquantel sensitivity to a mammalian Ca^{2+} channel. *Int J Parasitol* 2003;33(12):1303–8.
- [41] Schena M, Shalon D, Davis RW, Brown PO. Quantitative monitoring of gene expression patterns with a complementary DNA microarray. *Science* 1995;270(5235):467–70.
- [42] Eisen MB, Spellman PT, Brown PO, Botstein D. Cluster analysis and display of genome-wide expression patterns. *Proc Natl Acad Sci USA* 1998;95(25):14863–8.
- [43] Hoffmann KF, Wynn TA, Dunne DW. Cytokine-mediated host responses during schistosome infections; walking the fine line between immunological control and immunopathology. *Adv Parasitol* 2002;52:265–307.
- [44] Basch PF. Why do schistosomes have separate sexes? *Parasitol Today* 1990;6(5):160–3.
- [45] Ribeiro-Paes JT, Rodrigues V. Sex determination and female reproductive development in the genus *Schistosoma*: a review. *Rev Inst Med Trop Sao Paulo* 1997;39(6):337–44.
- [46] Gupta BC, Basch PF. The role of *Schistosoma mansoni* males in feeding and development of female worms. *J Parasitol* 1987;73(3):481–6.
- [47] Basch PF. *Schistosomes: development, reproduction, and host relations*. New York: Oxford University Press Inc.; 1991.
- [48] Robinson DL. Egg-laying by *Schistosoma mansoni* in vitro. *Ann Trop Med Parasitol* 1960;54:112–7.
- [49] Popiel I, Basch PF. Reproductive development of female *Schistosoma mansoni* (Digenea: Schistosomatidae) following bisexual pairing of worms and worm segments. *J Exp Zool* 1984;232(1):141–50.
- [50] Kohn AB, Anderson PA, Roberts-Misterly JM, Greenberg RM. Schistosome calcium channel beta subunits. Unusual modulatory effects and potential role in the action of the antischistosomal drug praziquantel. *J Biol Chem* 2001;276(40):36873–6.
- [51] Kohn AB, Lea J, Roberts-Misterly JM, Anderson PA, Greenberg RM. Structure of three high voltage-activated calcium channel alpha1 subunits from *Schistosoma mansoni*. *Parasitology* 2001;123(Part 5):489–97.
- [52] Andrews P. Praziquantel: mechanisms of anti-schistosomal activity. *Pharmacol Ther* 1985;29(1):129–56.
- [53] Redman CA, Robertson A, Fallon PG, et al. Praziquantel: an urgent and exciting challenge. *Parasitol Today* 1996;12(1):14–20.
- [54] Pica-Mattoccia L, Cioli D. Sex- and stage-related sensitivity of *Schistosoma mansoni* to in vivo and in vitro praziquantel treatment. *Int J Parasitol* 2004;34(4):527–33.
- [55] Hashimoto T, Amagai M, Parry DA, et al. Desmoyokin, a 680 kDa keratinocyte plasma membrane-associated protein, is homologous to the protein encoded by human gene AHNK. *J Cell Sci* 1993;105(Part 2):275–86.
- [56] Haase H, Pagel I, Khalina Y, et al. The carboxyl-terminal ahnak domain induces actin bundling and stabilizes muscle contraction. *FASEB J* 2004;18(7):839–41.
- [57] Siddiqui AA, Zhou Y, Podesta RB, et al. Characterization of Ca^{2+} -dependent neutral protease (calpain) from human blood flukes, *Schistosoma mansoni*. *Biochim Biophys Acta* 1993;1181(1):37–44.
- [58] Thompson DP, Chen GZ, Sample AK, Semeyn DR, Bennett JL. Calmodulin: biochemical, physiological, and morphological effects on *Schistosoma mansoni*. *Am J Physiol* 1992;251(6 Part 2):R1051–8.
- [59] Stewart TJ, Smith AL, Havercroft JC. Analysis of the complete sequence of a muscle calcium-binding protein of *Schistosoma mansoni*. *Parasitology* 1992;105(Part 3):399–408.
- [60] Silva CL, Cunha VM, Mendonca-Silva DL, Noel F. Evidence for ryanodine receptors in *Schistosoma mansoni*. *Biochem Pharmacol* 1998;56(8):997–1003.
- [61] Goodman DS, Deykin D, Shiratori T. The formation of cholesterol esters with rat liver enzymes. *J Biol Chem* 1964;239:1335–45.
- [62] Cases S, Novak S, Zheng YW, et al. ACAT-2, a second mammalian acyl-CoA:cholesterol acyltransferase. Its cloning, expression, and characterization. *J Biol Chem* 1998;273(41):26755–64.
- [63] Cases S, Smith SJ, Zheng YW, et al. Identification of a gene encoding an acyl CoA:diacylglycerol acyltransferase, a key enzyme in triacylglycerol synthesis. *Proc Natl Acad Sci USA* 1998;95(22):13018–23.
- [64] Weiss SB, Kennedy EP, Kiyasu JY. The enzymatic synthesis of triglycerides. *J Biol Chem* 1960;235:40–4.
- [65] Cases S, Stone SJ, Zhou P, et al. Cloning of DGAT2, a second mammalian diacylglycerol acyltransferase, and related family members. *J Biol Chem* 2001;276(42):38870–6.
- [66] Meyer F, Meyer H, Bueding E. Lipid metabolism in the parasitic and free-living flatworms. *Schistosoma mansoni* and *Dugesia dorotocephala*. *Biochim Biophys Acta* 1970;210(2):257–66.
- [67] Barrett J. *Biochemistry of parasitic helminths*. London: MacMillan; 1981.
- [68] Lawrence JD. The ingestion of red blood cells by *Schistosoma mansoni*. *J Parasitol* 1973;59(1):60–3.

- [69] Brindley PJ, Kalinna BH, Dalton JP, et al. Proteolytic degradation of host hemoglobin by schistosomes. *Mol Biochem Parasitol* 1997;89(1):1–9.
- [70] Smyth D, McManus DP, Smout MJ, et al. Isolation of cDNAs encoding secreted and transmembrane proteins from *Schistosoma mansoni* by a signal sequence trap method. *Infect Immun* 2003;71(5):2548–54.
- [71] Skelly PJ, Da'dara A, Harn DA. Suppression of cathepsin B expression in *Schistosoma mansoni* by RNA interference. *Int J Parasitol* 2003;33(4):363–9.
- [72] Boyle JP, Wu XJ, Shoemaker CB, Yoshino TP. Using RNA interference to manipulate endogenous gene expression in *Schistosoma mansoni* sporocysts. *Mol Biochem Parasitol* 2003;128(2):205–15.
- [73] Ashton PD, Curwen RS, Wilson RA. Linking proteome and genome: how to identify parasite proteins. *Trends Parasitol* 2001;17(4):198–202.
- [74] Colland F, Jacq X, Trouplin V, et al. Functional proteomics mapping of a human signaling pathway. *Genome Res* 2004;14(7):1324–32.